



# Stress of cupric ion and oxytetracycline in *Chlorella vulgaris* cultured in swine wastewater

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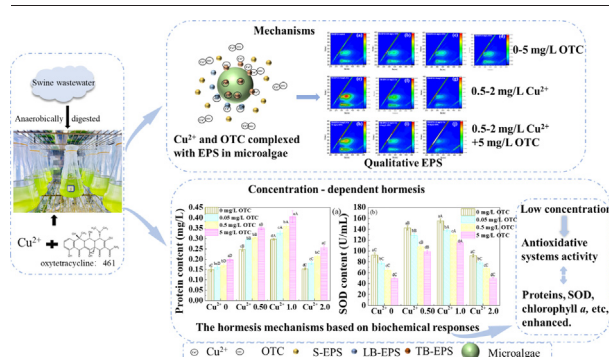
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## HIGHLIGHTS

- Either OTC or Cu<sup>2+</sup> induced hormesis in NH<sub>3</sub>-N removal via microalgae culturing.
- Lipid content and mass of microalgae maximized when cultured in 0.05 mg/L of OTC.
- Cupric ions induced the oxidative stress response of microalgae in combined stress.
- OTC mitigated the toxicity of Cu<sup>2+</sup> on microalgae under the combined stress.

## GRAPHICAL ABSTRACT



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## ABSTRACT

*Chlorella* culturing has the advantages in treatment of wastewater including swine wastewater from anaerobic digesters due to the product of biolipids and the uptake of carbon dioxide. However, there often exist high concentrations of antibiotics and heavy metals in swine wastewater which could be toxic to *Chlorella* and harmful to the biological systems. This study examined the stress of cupric ion and oxytetracycline (OTC) at various concentrations on the nutrient removal and biomass growth in *Chlorella vulgaris* culturing in swine wastewater from anaerobic digesters, and its biochemical responses were also studied. Results showed that dynamic hormesis of either OTC concentration or cupric ion one on *Chlorella vulgaris* were confirmed separately, and the presence of OTC not only did not limit biomass growth and lipids content of *Chlorella vulgaris* but also could mitigate the toxicity of cupric ion on *Chlorella vulgaris* in combined stress of Cu<sup>2+</sup> and OTC. Extracellular polymeric substances (EPS) of *Chlorella vulgaris* were used to explain the mechanisms of stress for the first time. The content of proteins and carbohydrates in EPS increased, and the fluorescence spectrum intensity of tightly-bound EPS (TB-EPS) of *Chlorella vulgaris* decreased with increasing concentration of stress because Cu<sup>2+</sup> and OTC may be chelated with proteins of TB-EPS to form non-fluorescent characteristic chelates. The low concentration of Cu<sup>2+</sup> ( $\leq 1.0$  mg/L) could enhance the protein content and promote the activity of superoxide dismutase (SOD) while these parameters were decreased drastically under 2.0 mg/L of Cu<sup>2+</sup>. The activity of adenosine

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triphosphatase (ATPase) and glutathione (GSH) enhanced with the increase of OTC concentration under combined stress. This study helps to comprehend the impact mechanisms of stress on *Chlorella vulgaris* and provides a novel strategy to improve the stability of microalgae systems for wastewater treatment.

## 1. Introduction

Microalgae can effectively remove and recover nitrogen and phosphorus from various wastewater to synthesize intracellular proteins, lipids, pigments, nucleic acids, and ATPase, particularly in swine wastewater, (Luo et al., 2016; Li et al., 2018; Li et al., 2020b). Microalgae also assimilate CO<sub>2</sub> as inorganic carbon sources and release O<sub>2</sub> to bacteria based on microalgal-bacterial consortium systems, which maximumly removed exceeded 90 % of CO<sub>2</sub> and harvest high concentrations of biomass in high-rate algal ponds (HRAP) (Li et al., 2022d; Serejo et al., 2015; Bahr et al., 2014; Li et al., 2022b).

However, there are abundant heavy metals and antibiotics in swine wastewater, which affected the performances of biological processes for the treatment. Since heavy metals such as Cu<sup>2+</sup> and Zn<sup>2+</sup> are also often used as feed additives, >80 % of which are excreted with swine manures and Cu<sup>2+</sup> at 135–374 mg/kg dry-manure and at 40 mg/L in swine wastewater and Zn<sup>2+</sup> was detected at 15 mg /L (Suzuki et al., 2010; Cestonaro do Amaral et al., 2014). Various antibiotics also are commonly used to prevent and treat pig diseases, and promote the growth of food animals as feed additives (Khan et al., 2008), which can be present in livestock wastes. The highest concentration of 2.02 mg/L of OTC and 166.7 mg/kg of tetracyclines (TCs) was detected in swine wastewater and manure, respectively (Ben et al., 2013; Wang et al., 2016).

Heavy metals and antibiotics as stressors induce the hormesis of the biological processes in wastewater treatment expressing a dose-response relationship. At low concentrations, OTC is regarded as a nutrient carbon resource (Wu et al., 2022). Low concentrations of Cu<sup>2+</sup> and Zn<sup>2+</sup> can be used as micronutrients to participate material construction of microalgae in the intracellular (Christenson and Sims, 2011). For example, Zn<sup>2+</sup> can be used as cofactors in the synthesis of proteins and enzyme systems of microalgae cells (Zhou et al., 2018), and Cu<sup>2+</sup> and Fe are components of photosynthetic electronic operation proteins in microalgae photosynthesis (Li et al., 2020b; Miazek et al., 2015). However, Zn<sup>2+</sup> will affect microalgae for the absorption of Ca<sup>2+</sup> at high concentrations, resulting in the activity of ATPase reduction (Liu et al., 2021b). Cu<sup>2+</sup> replaces Mg<sup>2+</sup> in chlorophyll, affecting the composition of cytochrome and the transport of photosynthetic electronics, thereby causing the photosynthesis of microalgae to decrease (Xiao et al., 2023). Low concentrations stimulate the synthesis of antioxidant enzymes, antioxidants, protein, and chlorophyll (Xiao et al., 2022), but high concentrations of Cu<sup>2+</sup>, Zn<sup>2+</sup>, and antibiotics will cause the accumulation of microalgae reactive oxygen species (ROS) and destroy the balance of the antioxidant system (Nicodemus et al., 2020; Zhou et al., 2018).

Effects of high concentration of Cu<sup>2+</sup> on the biomass of microalgae and the capacity of nutrients removal and the hormesis induced by Cu<sup>2+</sup> stress were reported. Liu et al. (2021a) reported that the low concentrations of Cu<sup>2+</sup> promoted the biomass of *Desmodesmus* sp. CHX1, thus the removal efficiency of Cu<sup>2+</sup> by microalgae increased. However, when Cu<sup>2+</sup> concentrations reached 2–3 mg/L, the growth of microalgae began to be inhibited. Hamed et al. (2017) researched the growth and hormetic effects of two green microalgae in the presence of Cu<sup>2+</sup>, which induced membrane damage in microalgae, and proline, glutathione levels, the activity of glutathione reductase (GR) and SOD enhanced to mitigate oxidative stress caused by Cu stress. A study by Li et al. (2022b) found that the content of lipids increased when gamma-aminobutyric acid (GABA) combined with copper ions, and hormesis was observed. For example, Cu<sup>2+</sup> stress significantly up-regulated the content of SOD and peroxidase (POD) for eliminating ROS during the early stage of culturing, subsequently, the antioxidant enzymes activities declined with an increase of culture time. These studies

indicated that Cu<sup>2+</sup> stress could induce the hormesis of microalgae, resulting in performances of nutrients removal of the biological processes affected, and the parameters such as biomass, antioxidant enzymes, and reactive oxygen species in microalgae could be as test endpoints.

OTC belongs to bacteriostatic antibiotics and is one of the tetracyclines. Siedlewicz et al. (2020) reported that the photosynthesis system II of microalgae and cyanobacteria was disrupted, photosynthetic electronics transport was blocked, and photosynthetic pigment synthesis decreased under 8 µg/cm<sup>3</sup> of OTC. A study has reported that the treatment of OTC-containing swine wastewater with *Lemna aequinoctialis* enhanced the synthesis of protein and lutein at low concentrations, but on the contrary at high concentrations, led to increasing intracellular H<sub>2</sub>O<sub>2</sub> accumulation, inducing protein carbonylation, and restraining the synthesis of related enzymes in the vitamin C synthesis pathway (Hu et al., 2019; Hu et al., 2021). Nevertheless, single contaminants caused the hormesis of microalgae were widely studied, and whether heavy metals and antibiotics can have hormesis on the microalgae treatment process would be studied further.

Microalgae can remove antibiotics via biodegrading and bio-adsorbing (Michelon et al., 2022; Wu et al., 2022). Heavy metals are removed through biosorption and bioaccumulation of microalgae (Hu et al., 2020). However, few studies have reported on the removal mechanism by extracellular polymeric substances (EPS) of microalgae. EPS is a polymer with a variety of functions on the surface of cells, which is also a defense mechanism of microorganisms, and protects microbial cells from dehydration and toxic substances (Sheng et al., 2010; Wu et al., 2022b). The EPS of microorganisms is mainly composed of proteins and carbohydrates, which can adsorb organic and inorganic pollutants due to the binding sites (D'Abzac et al., 2010; Laspidou and Rittmann, 2002). EPS contains a large number of various types of functional groups of aromatic compounds and unsaturated fat chains, which have fluorescent properties, thus, the 3-dimensional excitation-emission matrix (3D-EEM) fluorescence spectrum is often used to study the physicochemical properties of EPS under stress (Sheng et al., 2010). In our study, we questioned whether the mechanisms of Cu<sup>2+</sup> and OTC on microalgae can be explored and speculated by qualitative and quantitative analysis of EPS.

There are few studies focused on the effects of multiple pollutions on *Chlorella vulgaris* growth and the capacity of wastewater treatment by *Chlorella vulgaris*. In this study, we aimed to reveal the effects of continuous and stable stress of OTC and Cu<sup>2+</sup> on microalgae processes in swine wastewater, the hormesis in microalgae, and the mechanisms of the stress acting on *Chlorella vulgaris*. The antioxidant stress response of *Chlorella vulgaris* was measured by analyzing the content of SOD, H<sub>2</sub>O<sub>2</sub>, GHS, and two ATPases. 3D-EEM of EPS and Fourier transform infrared spectrum (FTIR) of microalgae were used to reveal the mechanism of microalgae with OTC and Cu<sup>2+</sup>. Finally, the intermediates and biodegradation pathways of OTC were proposed.

## 2. Materials and methods

### 2.1. Microalgal strain

*Chlorella vulgaris* comes from Freshwater Algae Culture Collection at the Institute of Hydrobiology, FACHB, Wuhan, China, strain number was FACHB-10, and is cultured in a sterilized BG-11 medium. Table S1 and S2 are the BG-11 media components. The pH of the medium is maintained between 7.0–7.2 and then sterilized at 121 °C for 15 min. The temperature in the photobioreactor was 25 ± 1 °C, the light conditions were 1000–2000 Lux, and the day/night was 12:12 h.

## 2.2. Swine wastewater from anaerobic digesters

The experimental wastewater was taken from the anaerobic digester effluent of a swine farm in Hengyang, Hunan, which did not contain OTC and only included a level of  $\mu\text{g}$  of  $\text{Cu}^{2+}$ . The collected swine wastewater was statically settled, centrifuged, and filtered to remove suspended solids. The pretreated was refrigerated at 4 °C. The characteristics of swine wastewater were showed in Table S3.

## 2.3. Chemicals

$\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$  (Analytically Pure, Shanghai, China) was used as a source of 0.1 g/L of  $\text{Cu}^{2+}$  in swine wastewater, and oxytetracycline hydrochloride (High Purity Grade) as a source of 0.5 g/L of OTC was derived from Bomei Biotechnology Co., LTD, Hefei, China.

## 2.4. Experimental design

Three different concentrations of 0.50, 1.0, and 2.0 mg/L of  $\text{Cu}^{2+}$  were severally combined with three different concentrations of 0.05, 0.5, and 5 mg/L of OTC forming 16 experimental groups, which were set through literature research and pre-experiment. The pretreated swine wastewater was diluted and sterilized and *Chlorella vulgaris* was inoculated in 500 mL of Erlenmeyer flasks with 400 mL of swine wastewater for every experiment group, and the inoculation biomass of *Chlorella vulgaris* was controlled at  $\text{OD}_{680}$  0.1. The flasks were placed in a photobioreactor for 12 days of culturing. The culture conditions were consistent with Section 2.1. The flasks were shaken three times every day. And the concentrations of  $\text{Cu}^{2+}$  and OTC were supplemented every 24 h, to maintain the original concentration level in swine wastewater.

## 2.5. Chemical analysis

5 mL of solution inoculated algae was centrifuged and filtered for the determination of  $\text{Cu}^{2+}$  concentration by atomic absorption spectrometry (AAS, Agilent 3510, USA) by Li et al. (2023). The high-performance liquid chromatography (HPLC, Agilent, Palo alto, California, U.S.A.) by Lin et al. (2019) was used to measure OTC, which was detailedly described in supplementary materials.

*Chlorella vulgaris* biomass was determined by collecting 3 mL of solution using a visible light spectrophotometer (721, Sunny Hengping Scientific Instrument Co., Ltd. Shanghai) at 680 nm, and the dry weight and absorbance of *Chlorella vulgaris* biomass showed a certain linear relationship. Dry weight was calculated as follows equation:

$$\text{DW}(\text{g/L}) = 0.3225 \times \text{OD}_{680}, R^2 = 0.9970$$

Nessler's reagent colorimetric methods by Li et al. (2023) were used to analyze  $\text{NH}_3\text{-N}$ , and TP was analyzed using potassium persulfate digestion methods by Luo et al. (2016).

Lipids extraction from dry biomass of *Chlorella vulgaris* via the Gravimetric method by Zhang et al. (2021a). Microalgae were collected and freeze-dried after 12 days of culturing. 0.1 g of dried biomass was weighed into a 25 mL of spiral tube with 10 mL of a methanol-chloroform mixture (methanol: chloroform = 1:2), and the tubes were ultrasound in an ultrasonic machine for 1 h under the maximum power. Microalgae were extracted overnight for 24 h in a shaker with shaking at 130 rpm, 27 °C. Filtering the extract into a pre-weighed glass tube with a 0.22  $\mu\text{m}$  filter, then 3 mL of ultrapure water was added, rapidly shaken for 1 min, and finally stood for 1 h to remove the upper aqueous phase. The tubes were evaporated at 50 °C in the oven. The change in the weight of the glass tube was the amount of lipids in the dry biomass of *Chlorella vulgaris*. The lipids content was expressed as lipid/dry weight (g/g).

The determination of chlorophyll *a* was determined by Li et al. (2020a). The content of protein and the activity of superoxide dismutase (SOD), glutathione (GSH), hydrogen peroxide ( $\text{H}_2\text{O}_2$ ),  $\text{Ca}^{++}\text{Mg}^{++}$ -ATPase and

$\text{Na}^{++}\text{K}^{++}$ -ATPase were determined by ELISA kit (Shenzhen, China) with 0.1 g microalgae respectively.

## 2.6. Data analysis

All experimental groups were repeated three times, with results expressed as mean  $\pm$  standard error ( $n = 3$ ). IBM SPSS Statistics 26 was used for statistical analyses of data. One-way analysis of variance (ANOVA) and two-way ANOVA were used to determine the significant difference of these parameters under stress. A value of  $p < 0.05$  represents the significant difference. All charts were graphed by Origin. Chem Draw 20.0 was used to draw the molecular structures.

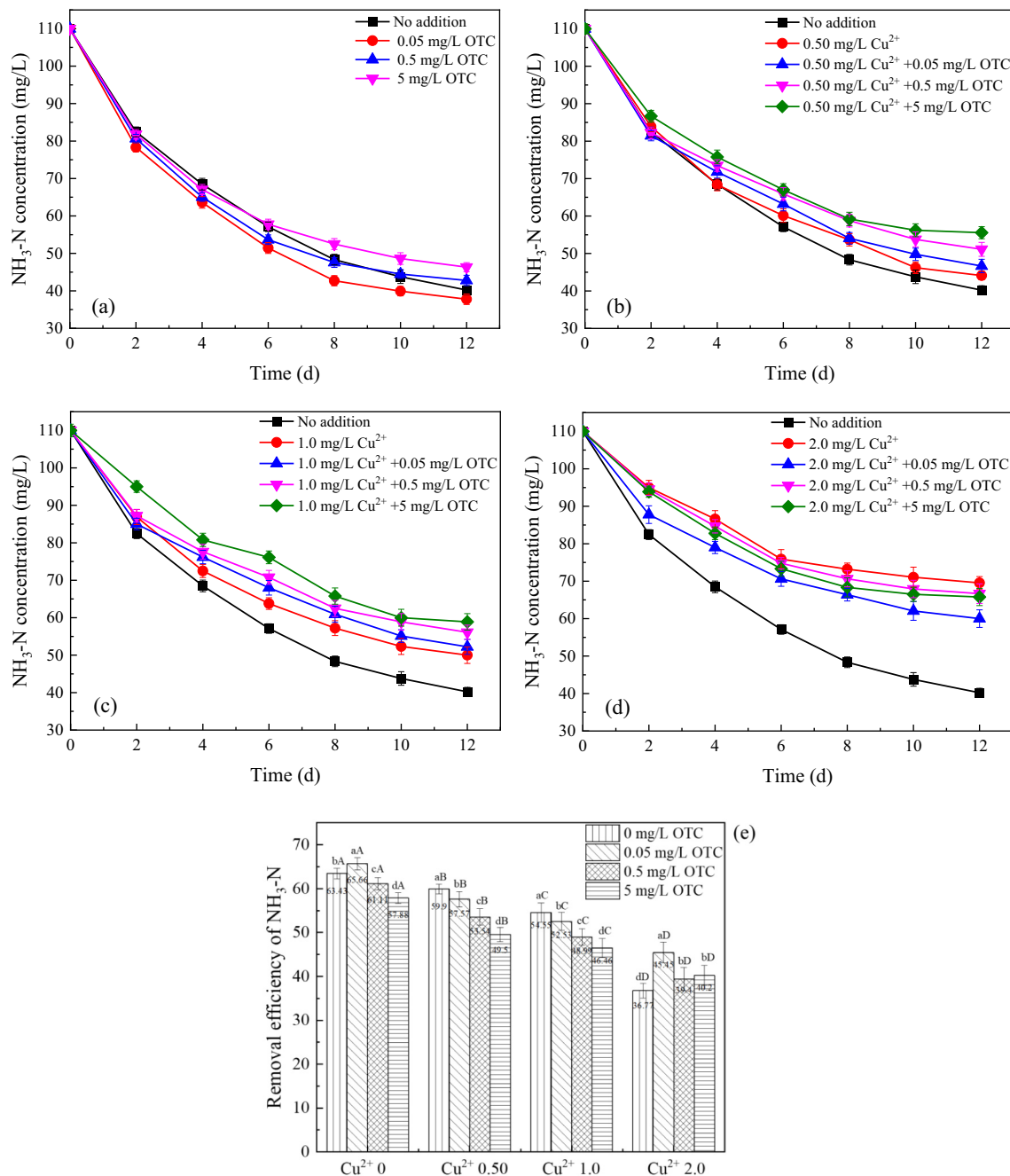
## 3. Results and discussion

### 3.1. $\text{NH}_3\text{-N}$ removal by *Chlorella vulgaris* under stress

$\text{NH}_3\text{-N}$  is the preferred form of nitrogen utilization by microalgae. The stress of  $\text{Cu}^{2+}$  and OTC on *Chlorella vulgaris* in the removal of  $\text{NH}_3\text{-N}$  in swine wastewater treatment was followed in Fig. 1. Fig. 1a and e showed the removal efficiency of  $\text{NH}_3\text{-N}$  reached 63.43 %, 65.66 %, 61.11 %, and 57.88 % respectively in the control group and single OTC. It found that in the presence of 0.05 mg/L of OTC, the removal efficiency of  $\text{NH}_3\text{-N}$  was slightly improved in swine wastewater, and within 6–8 days of *Chlorella vulgaris* culturing, the removal of  $\text{NH}_3\text{-N}$  also was promoted in the single high concentrations of OTC while was reduced subsequently. This result was similar to the growth of *Chlorella vulgaris* and the study by Oliveira et al. (2023). The treatment performance of  $\text{NH}_3\text{-N}$  was optimal by microalgae under the appropriate conditions, which provided the best condition for microalgae growth. It is described as "hormesis". The sub-toxicity of low concentrations stimulates but inhibits at high concentrations in organisms (Erofeeva, 2022). Hormesis is a highly consistent dose-time-response relationship (Agathokleous and Calabrese, 2022). At a low concentration, biomass growth of *Chlorella vulgaris* increased compared to the control group, which could correspondingly promote the removal efficiency of  $\text{NH}_3\text{-N}$ . However, when exceeding adequate concentration, the growth of microalgae began to be inhibited with the culture time increasing. When the single  $\text{Cu}^{2+}$  was present in Fig. 1b, c, and d, the removal efficiency of  $\text{NH}_3\text{-N}$  was 59.90 %, 54.55 %, and 36.77 %, respectively. The ability of  $\text{NH}_3\text{-N}$  removal by *Chlorella vulgaris* was greatly affected by  $\text{Cu}^{2+}$  stress. The removal efficiency of  $\text{NH}_3\text{-N}$  was further reduced when  $\text{Cu}^{2+}$  was combined with OTC. This may be because the content of glutamine synthetase (GS) decreased under combined stress (Fig. S1), which can catalyze glutamic acid and ammonium to synthesize glutamine (Gln) and is also an index for measuring the assimilation level of ammonia nitrogen (Qin et al., 2022). Results showed the hormesis of OTC concentration on *Chlorella vulgaris* in  $\text{NH}_3\text{-N}$  removal processes. The maximum removal efficiency of  $\text{NH}_3\text{-N}$  by *Chlorella vulgaris* was only 65.66 % and its removal ability was slightly inferior to other algae in swine wastewater. This may be the ammonia inhibition due to high initial  $\text{NH}_3\text{-N}$  concentration, which would decrease the growth rate of *Chlorella vulgaris*, and the magnitude of this difference in inhibition is related to our microalgae species (Xia and Murphy, 2016). Higher concentrations of  $\text{NH}_3\text{-N}$  may be toxic to *Chlorella vulgaris*.

### 3.2. The changes of $\text{Cu}^{2+}$ and OTC concentrations in swine wastewater

In order to better study the stress of  $\text{Cu}^{2+}$  and OTC on microalgae, the trend of the two pollutants in swine wastewater was monitored. Fig. 2 showed that the concentration of  $\text{Cu}^{2+}$  declined significantly within 2 h, mainly because  $\text{Cu}^{2+}$  was adsorbed by *Chlorella vulgaris*. The trend of  $\text{Cu}^{2+}$  concentration was similar to single  $\text{Cu}^{2+}$  when combined with 0.05 mg/L of OTC (Fig. 2a, b, and c). However, combined with the 0.5 and 5 mg/L of OTC, the  $\text{Cu}^{2+}$  concentration decreased obviously, which was most pronounced in the group of 2.0 mg/L of  $\text{Cu}^{2+}$  and 5 mg/L of OTC (Fig. 2c). In all the groups, the concentration of OTC decreased



**Fig. 1.** Stress of different concentrations of  $\text{Cu}^{2+}$  and OTC on  $\text{NH}_3\text{-N}$  concentration during 12 days of *Chlorella vulgaris* culturing in swine wastewater. (a) 0 mg/L of  $\text{Cu}^{2+}$ ; (b) 0.50 mg/L of  $\text{Cu}^{2+}$ ; (c) 1.0 mg/L of  $\text{Cu}^{2+}$ ; (d) 2.0 mg/L of  $\text{Cu}^{2+}$ . (e) The removal efficiency of  $\text{NH}_3\text{-N}$ . Error bars are expressed as standard deviation ( $n = 3$ ). Different letters represent that they are different significantly ( $p < 0.05$ , lowercases represent the same concentration of  $\text{Cu}^{2+}$ , and uppercases represent the same concentration of OTC).

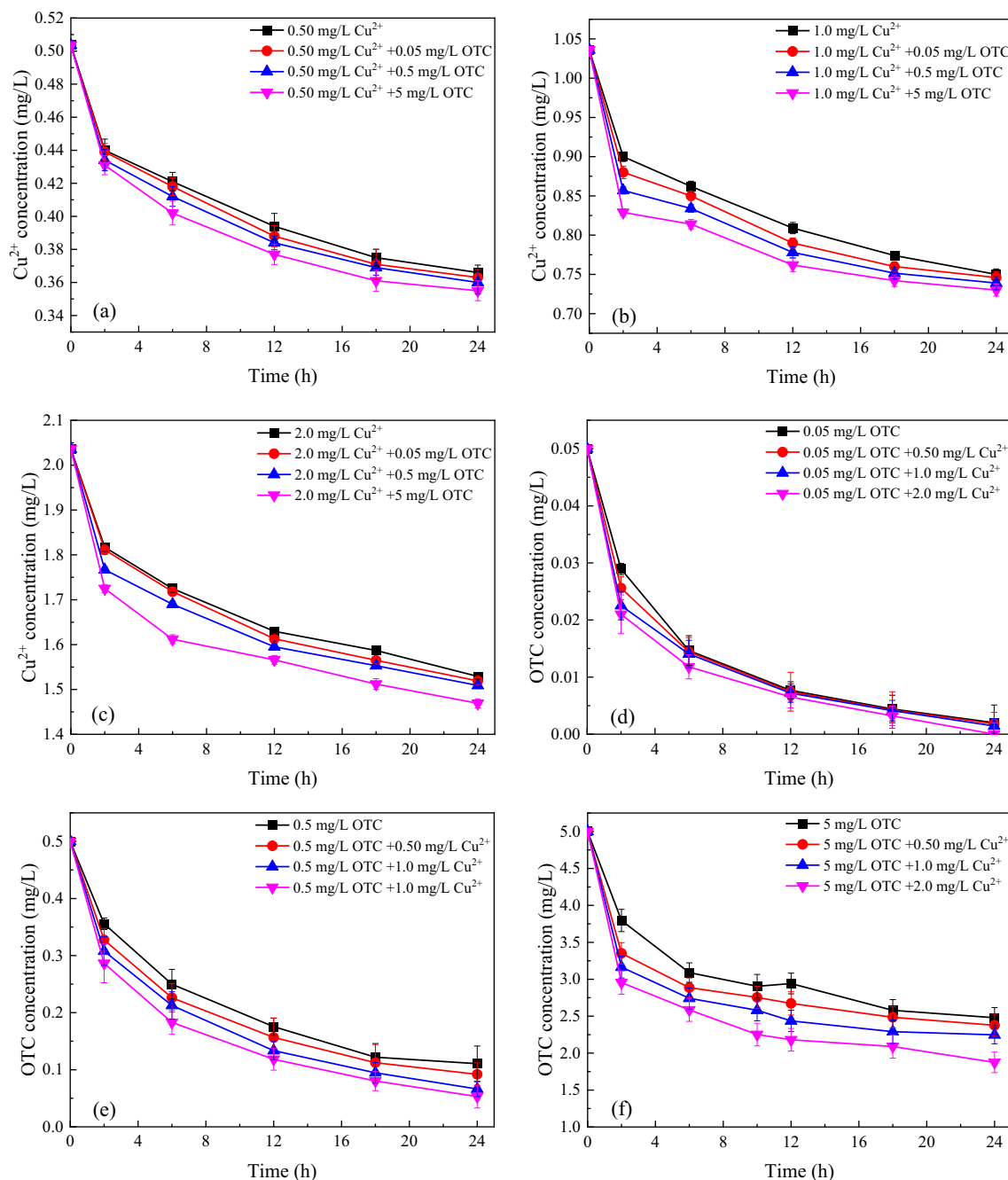
significantly within 2 h, mainly because of the biodegradation by *Chlorella vulgaris* (Wu et al., 2022). In all the groups of 0.05 mg/L of OTC, the concentration of OTC decreased by >96 % within 24 h (Fig. 2d). Fig. 2e and f showed the higher the concentration of  $\text{Cu}^{2+}$ , the more OTC concentration decreased, which was similar to the dynamic change of  $\text{Cu}^{2+}$  mentioned previously. But the decline rate of OTC in swine wastewater slowed down with the increasing culture time, especially in the group of 5 mg/L of OTC the concentration of OTC varied little after 12 h. Results indicated that the change of OTC concentration was greatly affected by  $\text{Cu}^{2+}$ , which changed significantly at the high concentration of  $\text{Cu}^{2+}$ . This may be because  $\text{Cu}^{2+}$  have a large potential of complexation affinities with antibiotics including organic ligands, which promotes the adsorption of

antibiotics by microalgae through  $\text{Cu}^{2+}$  bridging effect, similarly, antibiotics can enhance the internalization of  $\text{Cu}^{2+}$  (Zhao et al., 2013; You et al., 2022).

### 3.3. Biomass growth of *Chlorella vulgaris* under stress

Fig. 3a showed during the first 6 days of culture, the presence of OTC promoted the growth of *Chlorella vulgaris* compared to the control group (0.331 g/L). After the sixth day, *Chlorella vulgaris* growth was gradually inhibited in the single OTC of 0.5 and 5 mg/L, and was about 0.312 and 0.294 g/L, respectively. Interestingly, in 0.05 mg/L of OTC, the biomass of *Chlorella vulgaris* was still higher than the control group, and the final

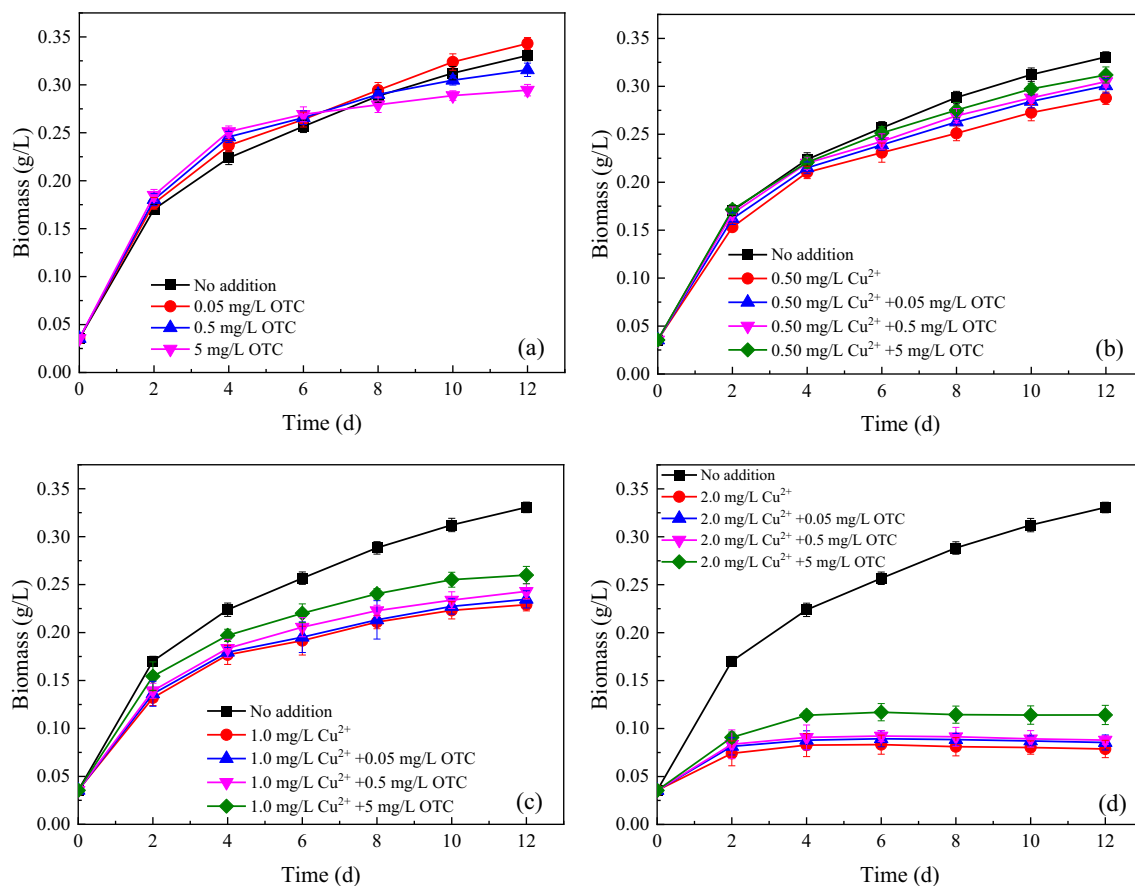




**Fig. 2.** Changes of  $\text{Cu}^{2+}$  and OTC concentration at stress of different concentrations of  $\text{Cu}^{2+}$  and OTC within 24 h of *Chlorella vulgaris* culturing in swine wastewater. (a) 0.50 mg/L of  $\text{Cu}^{2+}$ ; (b) 1.0 mg/L of  $\text{Cu}^{2+}$ ; (c) 2.0 mg/L of  $\text{Cu}^{2+}$ ; (d) 0.05 mg/L of OTC; (e) 0.5 mg/L of OTC; (f) 5 mg/L of OTC. Error bars are expressed as standard deviation (n = 3).

biomass was 0.343 g/L. In Fig. 3b, c, and d, *Chlorella vulgaris* biomass reached 0.288, 0.229, and 0.0780 g/L, respectively, the presence of  $\text{Cu}^{2+}$  significantly inhibited the growth of *Chlorella vulgaris* biomass. These were similar with previous study. Microalgae were observed significantly hormesis of concentration-dependent growth in the presence of antibiotics and  $\text{Cu}^{2+}$  could exert greater inhibition on growth (You et al., 2022). OTC may be regarded as a carbon source for *Chlorella vulgaris* at low concentrations which simulated microalgae biomass (Wu et al., 2022). However, in the presence of 0.5 and 5 mg/L of OTC, these revealed the hormesis of concentration-time-dependent relationship. Fig. 3b showed that OTC presented, the inhibition of  $\text{Cu}^{2+}$  on biomass growth was alleviated, and the biomass yield of *Chlorella vulgaris* maximumly increased by 7.60 % compared with the single  $\text{Cu}^{2+}$ . The same results were more obvious in

Fig. 3c and d. This consistent with the changes of  $\text{Cu}^{2+}$  and OTC concentrations. In the group of 2.0 mg/L of  $\text{Cu}^{2+}$  (Fig. 3d), microalgae growth almost stopped from the 4th day, indicating that this concentration of  $\text{Cu}^{2+}$  expressed greater toxicity on microalgae, and the toxicity was significantly alleviated when 5 mg/L of OTC was added. The inhibition growth induced by  $\text{Cu}^{2+}$  was alleviated when combined with OTC. This may be because  $\text{Cu}^{2+}$  firstly complexed with OTC in swine wastewater, when OTC presented, and this detoxification effect was promoted with the concentration of OTC increased, reducing the direct effect of  $\text{Cu}^{2+}$  on *Chlorella vulgaris* (Wu and He, 2019). Nevertheless, exceeding 2.0 mg/L of  $\text{Cu}^{2+}$  caused severe and irreversible oxidative stress to microalgae, and combined with OTC to decrease the toxicity on *Chlorella vulgaris* become insignificant.



**Fig. 3.** Stress of different concentrations of  $\text{Cu}^{2+}$  and OTC on biomass growth during 12 days of *Chlorella vulgaris* culturing in swine wastewater. (a) 0 mg/L of  $\text{Cu}^{2+}$ ; (b) 0.50 mg/L of  $\text{Cu}^{2+}$ ; (c) 1.0 mg/L of  $\text{Cu}^{2+}$ ; (d) 2.0 mg/L of  $\text{Cu}^{2+}$ . Error bars are expressed as standard deviation ( $n = 3$ ).

### 3.4. The content change of protein, lipids, and chlorophyll in *Chlorella vulgaris* under stress

#### 3.4.1. Protein content

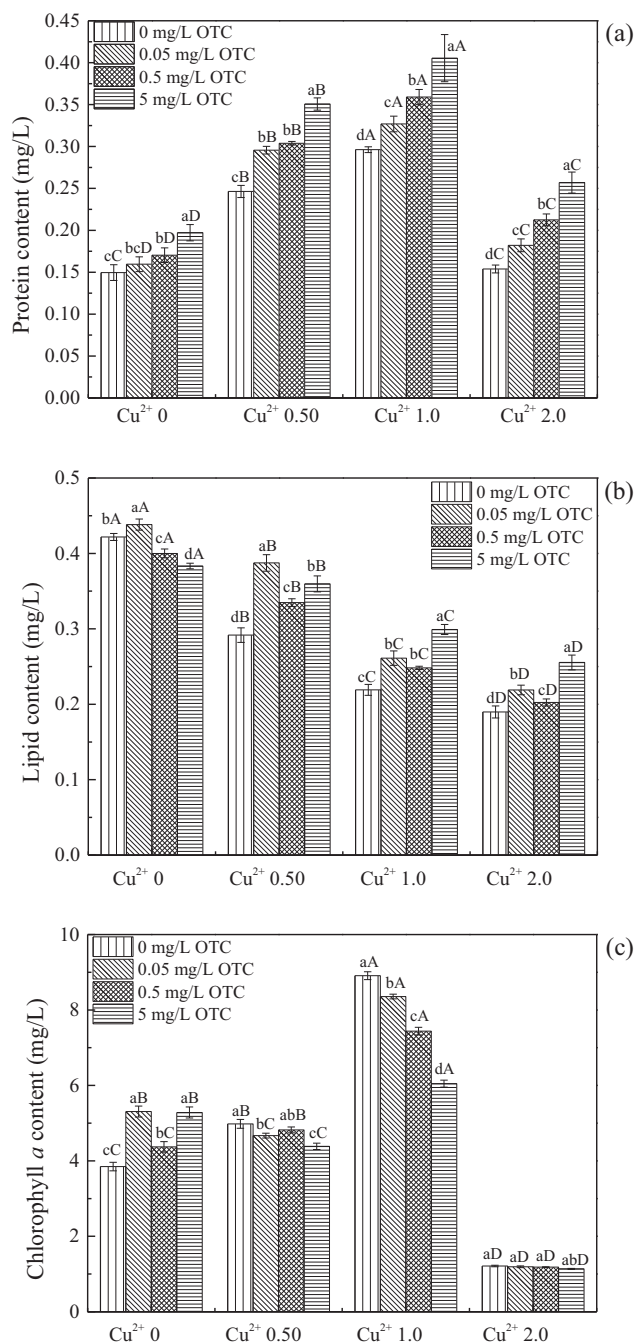
Protein content in microalgae plays an important role in analyzing the hormesis of *Chlorella vulgaris* under stress, including the sum of intracellular proteins and various enzymes. The sub-toxicity at low concentration will stimulate the synthesis of relevant protein to protect the cells from oxidative damage and promote the tolerance of virulence but adverse impact at high concentrations. Fig. 4a showed in low concentrations group of 0.50 and 1.0 mg/L of  $\text{Cu}^{2+}$ , protein content reached 0.246 and 0.296 g/g, which further increased to 0.351 and 0.405 g/g respectively when 5 mg/L of OTC was added. It showed that long-term exposure of constant low concentrations could induce the hormesis of microalgae, and stimulated the content increase of protein including corresponding plant chelates, metallothionein (Perales-Vela et al., 2006), and some oxidative stress enzymes (Sabatini et al., 2009) for maintaining the *Chlorella vulgaris* growth. Protein content decreased drastically in the group of 2.0 mg/L of  $\text{Cu}^{2+}$  (Fig. 4a), but similarly, when the concentration of OTC increased, the protein content of microalgae also increased correspondingly. Study proved that  $\text{Cu}^{2+}$  induced the hormesis of microalgae by stimulation at low concentrations but inhibition at high concentrations, however, the single  $\text{Cu}^{2+}$  would cause the disorder of the proteomic. Fortunately, CTC combined with  $\text{Cu}^{2+}$  declined this effect and up-regulated protein, which decreased the toxicity on microalgae (You et al., 2022). OTC and CTC both are belong to the tetracyclines (TCs), thus, the addition of OTC could increase the content of protein and slow the toxicity on *Chlorella vulgaris*. These were similar with the study by (Lu et al., 2015).

Changes in microalgae were induced by exposing to environmental stress such as the content of protein, chlorophyll, and antioxidant enzymes

to overcome stress. Unfortunately, excessive concentrations exceeded the tolerance of *Chlorella vulgaris*, and the balance of system of intramolecular was disrupted, resulting ROS accumulating in microalgae, membrane permeability enhancing, and structural and functional proteins being denaturalized and lost (Demidchik et al., 2014; You et al., 2022). Hormesis of microorganism can induce the different levels of survival, resistance, and tolerance to respond higher levels of stressors (Agathokleous et al., 2022).

#### 3.4.2. Lipids content

In Fig. 4b, In the control group, the lipids content of *Chlorella vulgaris* reached 0.423 g/g, which was higher than that of other types of *Chlorella vulgaris* in wastewater treatment (Daneshvar et al., 2018; Kumaran et al., 2023; Shen et al., 2015; Verma et al., 2022). Luo et al. (2016) treated anaerobic digestion of swine wastewater by *Coelastrella* sp., and the lipids content only accounted for 22.40–25.50 % of dry-weight cells. The high content of lipids made *Chlorella vulgaris* more potential as a raw material of biofuel. The study found that lipids reached a maximum of 0.438 g/g at 0.05 mg/L of OTC, indicating that low concentrations of OTC could not only promote the increase of biomass but also stimulate the accumulation of lipids. In a study by Xie et al. (2019), at concentrations of 1 mg/L of SMX and 5 mg/L of bisphenol A (BPA), the lipids content of *Chlamydomonas* sp. *Tai-0* reached the maximum (25–30 %), similarly, the lipids content gradually decreased when the pollutant concentration further increased. In the single concentrations of 0.50, 1.0, and 2.0 mg/L of  $\text{Cu}^{2+}$ , the content of lipids was 0.293, 0.219, and 0.190 g/g, respectively, and the lipids yield was inhibited by 55.08 % maximally. A similar result was reported by Martínez-Macias et al. (2019). This is because  $\text{Cu}^{2+}$  exerted negative effects on lipids content of microalgae. When  $\text{Cu}^{2+}$  and 5 mg/L OTC were presented simultaneously, the lipids content reached 0.387, 0.299, and 0.255 g/g, respectively, which increased by 32.53 %, 36.53 %, and



**Fig. 4.** Stress of different concentrations of  $\text{Cu}^{2+}$  and OTC on the content of protein, lipids, and chlorophyll *a* during 12 days of *Chlorella vulgaris* culturing in swine wastewater. (a) Protein content; (b) Lipids content; (c) Chlorophyll *a* content. Error bars are expressed as standard deviation ( $n = 3$ ). Different letters represent that they are different significantly ( $p < 0.05$ , lowercases represent the same concentration of  $\text{Cu}^{2+}$ , and uppercases represent the same concentration of OTC).

34.21 % compared with the single  $\text{Cu}^{2+}$ . The conclusion is similar with Li et al. (2022b). The reason is that combined stress of  $\text{Cu}^{2+}$  and OTC alleviated the toxicity and up-regulated relevant protein modification and expression, besides, the microalgal glycolysis pathway and the tricarboxylic acid cycle were accelerated, in which the content of ATPase of *Chlorella vulgaris* (Fig. 5d and e), and enzymes related to lipid production (acetyl-Co-enzyme carboxylase (ACCase)) were up-regulated, which promote glycogen decomposition and generate related fatty acids (Wang et al., 2021; Xue et al., 2017; Li et al., 2022a; Yang et al., 2023).

### 3.4.3. Chlorophyll *a* content

The photosynthetic pigment content of microalgae can be used as a measure of the photosynthetic rate. When microalgae grew under heavy metals stress,  $\text{Mg}^{2+}$  in chlorophyll molecules could be replaced by heavy metal ions, thereby destroying the chloroplast structure and affecting the photosynthesis process of microalgae (Bechaieb et al., 2016; Xiao et al., 2023).  $\text{Cu}^{2+}$  is a component of the photosynthetic electronics transporter of microalgae cells, and an appropriate amount of  $\text{Cu}^{2+}$  was present, which contributed to promoting the synthesis of chlorophyll *a* (Cheng et al., 2019). Compared with other metal ions (such as zinc ions),  $\text{Cu}^{2+}$  is easier to replace  $\text{Mg}^{2+}$  in molecules to form copper chlorophyll. What's more, a large number of ROS and  $\text{H}_2\text{O}_2$  will be accumulated during microalgae photosynthesis under stress, and they cannot be eliminated in chloroplasts in time, the production of chlorophyll will be inhibited (Edreva, 2005).

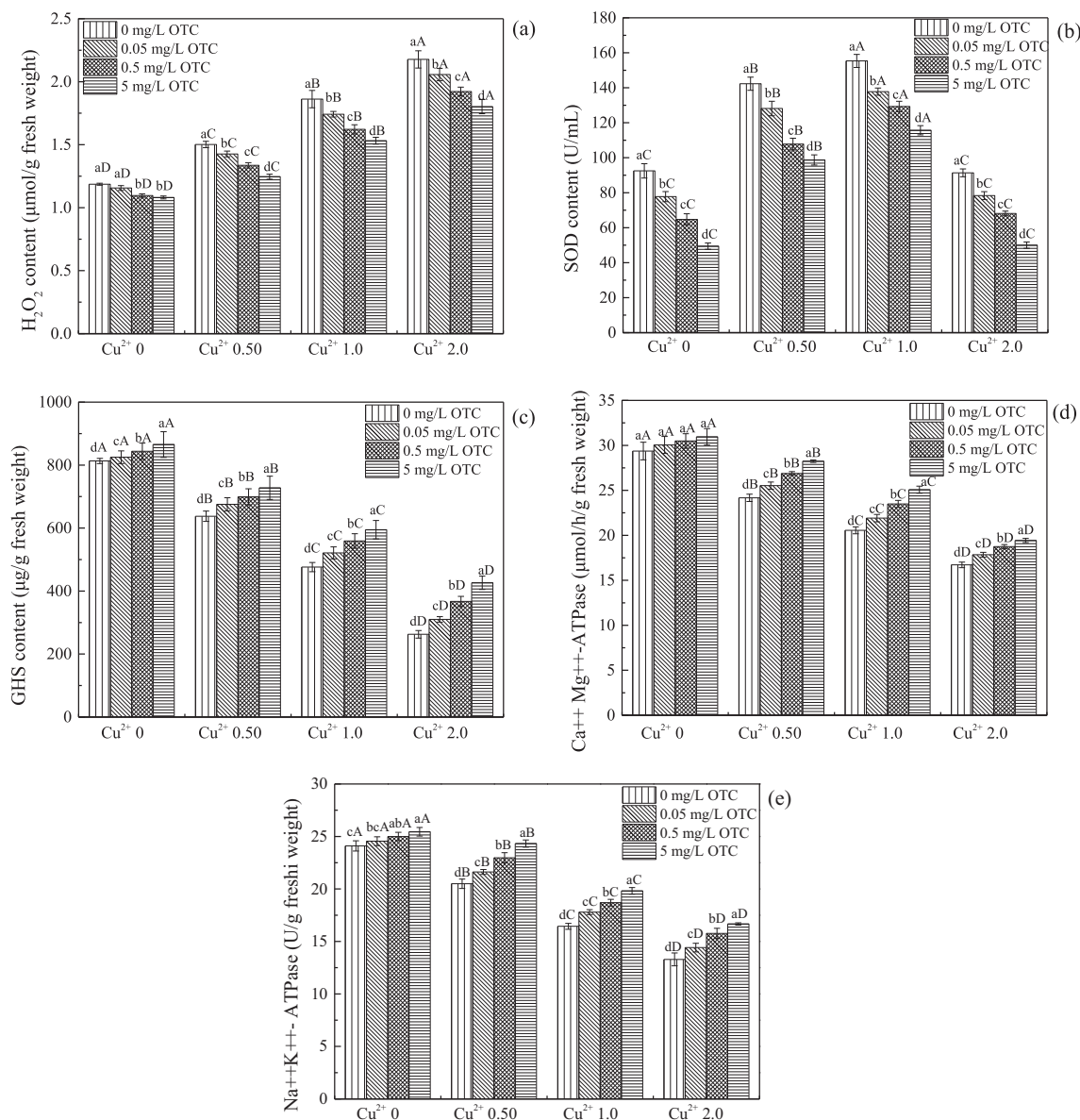
Fig. 4c showed that at the single OTC, the chlorophyll *a* content increased slightly to 5.31 mg/g. The single 0.50 and 1.0 mg/L of  $\text{Cu}^{2+}$  promoted the content of chlorophyll *a* in *Chlorella vulgaris*, and the content was about 4.98 and 8.91 mg/g, respectively., on the contrary, the chlorophyll *a* content decreased by 68.57 % in the single 2.0 mg/L of  $\text{Cu}^{2+}$ . The significant hormesis of microalgae in the aspect of chlorophyll *a* production induced by exposing to  $\text{Cu}^{2+}$  stress. The low concentrations of  $\text{Cu}^{2+}$  enhanced the synthesis of chlorophyll *a* and promoted photosynthetic efficiency. When the concentration of  $\text{Cu}^{2+}$  was excessive,  $\text{H}_2\text{O}_2$  was accumulated in *Chlorella vulgaris* cells and broke the normal biological function in microalgae (Zhang et al., 2020). When  $\text{Cu}^{2+}$  was combined with OTC, the chlorophyll *a* content decreased significantly ( $p < 0.05$ ). The combined stress of  $\text{Cu}^{2+}$  and OTC caused lipid peroxidation of the chloroplast membrane, and the synthesis pathway of chlorophyll *a* was intercepted (Chu et al., 2023).

### 3.5. Antioxidant stress response under stress

Hormesis is explained protective mechanisms of microalgae, is an over-compensation in oxidative stress response at low concentrations but oxidative damage under excessive stress, and is also the balance of ROS and the synthesis and decomposition of antioxidant enzymes and non-enzymatic antioxidants. Antioxidant enzymes could be as test endpoints of hormesis. (Erofeeva, 2022; Li et al., 2023).

$\text{H}_2\text{O}_2$  is a type of ROS, and its content can measure the level of oxidative damage of microalgae under stress (Danouche et al., 2020). The change of  $\text{H}_2\text{O}_2$  content was consistent with the results of biomass growth of *Chlorella vulgaris*. In Fig. 5a, 0.05–5 mg/L of OTC did not cause  $\text{H}_2\text{O}_2$  accumulation, and *Chlorella vulgaris* could regard OTC as a nutrient carbon source for absorption and utilization. The  $\text{H}_2\text{O}_2$  content was 1.19  $\mu\text{mol/g}$  fresh weight in the control group, and with the concentration increase of the single  $\text{Cu}^{2+}$ , the content of  $\text{H}_2\text{O}_2$  increased. The hormetic effect induced by adequate  $\text{Cu}^{2+}$  stimulated the accumulation of  $\text{H}_2\text{O}_2$ , which was eliminated by antioxidant enzymes. At 2.0 mg/L of  $\text{Cu}^{2+}$ , the content reached 2.18  $\mu\text{mol/g}$  fresh weight, and the cumulative of  $\text{H}_2\text{O}_2$  increased by 83.19 % compared with the control group. In the high environmental stress, a large of  $\text{H}_2\text{O}_2$  generated the balance of the synthesis and decomposition of enzymes being destroyed, resulting in peroxidative damage of microalgae, and the overcompensation became insignificant. It also was proved that  $\text{Cu}^{2+}$  plays a dominant role in stress of  $\text{Cu}^{2+}$  and OTC again. Less  $\text{H}_2\text{O}_2$  was accumulated in combined stress compared with the single  $\text{Cu}^{2+}$  groups. The reason is that  $\text{Cu}^{2+}$  complexed with OTC in swine wastewater, besides, combined stress of  $\text{Cu}^{2+}$  and OTC could decrease the toxicity on *Chlorella vulgaris*. The variation in  $\text{H}_2\text{O}_2$  content was similar to Danouche et al. (2020) and Chen et al. (2022).

SOD is the first defensive line of the antioxidant defense system in microalgae, catalyze superoxide anions to produce  $\text{H}_2\text{O}_2$  and  $\text{O}_2$ , and  $\text{H}_2\text{O}_2$  is then catalyzed by other enzymes to  $\text{H}_2\text{O}$  and  $\text{O}_2$  to prevent microalgae from oxidative damage (Mishra et al., 2021). Fig. 5b showed that the activity of SOD increased with the increase of  $\text{Cu}^{2+}$  concentrations, subsequently, it subsequently decreased drastically under 2.0 mg/L of OTC.



**Fig. 5.** Stress of different concentrations of  $\text{Cu}^{2+}$  and OTC on antioxidant stress response of *Chlorella vulgaris* during 7 days culturing in swine wastewater. (a)  $\text{H}_2\text{O}_2$  content; (b) SOD content; (c) GSH content; (d)  $\text{Ca}^{++}\text{Mg}^{++}$ -ATPase activity; (e)  $\text{Na}^{++}\text{K}^{++}$ -ATPase activity. Error bars are expressed as standard deviation (n = 3). Different letters represent that they are different significantly (p < 0.05, lowercases represent the same concentration of  $\text{Cu}^{2+}$ , and uppercases represent the same concentration of OTC).

At low concentrations, the antioxidant enzymes were activated for eliminating ROS, while excessive stress induced SOD activities decreasing. This was similar with the study reported by Cheng et al. (2018). Environmental stress would induce the activity of antioxidant defense system, however, excessive stress levels damaged severely the balance of system. Under combined stress of  $\text{Cu}^{2+}$  and OTC a declining activity of SOD compared with the single  $\text{Cu}^{2+}$  was observed. This is because  $\text{Cu}^{2+}$  combined with OTC, a declining toxic of  $\text{Cu}^{2+}$  impacted on *Chlorella vulgaris*, which decreased the damage induced by  $\text{Cu}^{2+}$  to the antioxidant system of microalgae. Thus, it could be speculated that the hormesis caused by combined stress would promote the tolerance and resistance of microalgae to environmental stress.

Glutathione (GSH) is a non-enzymatic water-soluble antioxidant in microalgae containing a group of sulfhydryl (-SH) that is complexed with heavy metals (Noctor et al., 1998). GSH can eliminate free radicals of ROS, even directly counteract some ROS, or act as a cofactor and substrate in enzymatic reactions to control ROS levels (Okamoto et al., 2001). For example, at high levels of ROS under environmental stress, with GR enzymes

catalyzing, GSH was converted to Glutathione (Oxidized) (GSSG) in the cytoplasm (Upadhyay et al., 2016). Fig. 5c showed that in the presence of the single  $\text{Cu}^{2+}$ , the content of GSH gradually decreased, and it decreased by 67.70 % in the group of 2.0 mg/L of  $\text{Cu}^{2+}$ , but in stress of  $\text{Cu}^{2+}$  and OTC, the consumption of GSH began to decrease. The reason is that the presence of OTC reduced the content of  $\text{Cu}^{2+}$  via bioaccumulation into the cells.

$\text{Ca}^{++}\text{Mg}^{++}$ -ATPase and  $\text{Na}^{++}\text{K}^{++}$ -ATPase catalyze ATP hydrolysis to produce ADP and inorganic phosphorus. The activity of ATPase is determined by the amount of inorganic phosphorus produced (Fig. 5d and e). In the single OTC, ATPase activity increased, because of the degrading of OTC by microalgae. With the single  $\text{Cu}^{2+}$  concentration rising, the ATPase activity was inhibited, but it gradually enhanced under combined stress of  $\text{Cu}^{2+}$  and OTC, especially in the group of  $\text{Cu}^{2+}$  and a high concentration of OTC. The reason is that combined stress up-regulated the modification and expression of proteins and various metabolic pathways, in which consumed a number of ATP. Under combined stress, the antioxidant capacity of microalgae was enhanced, and the



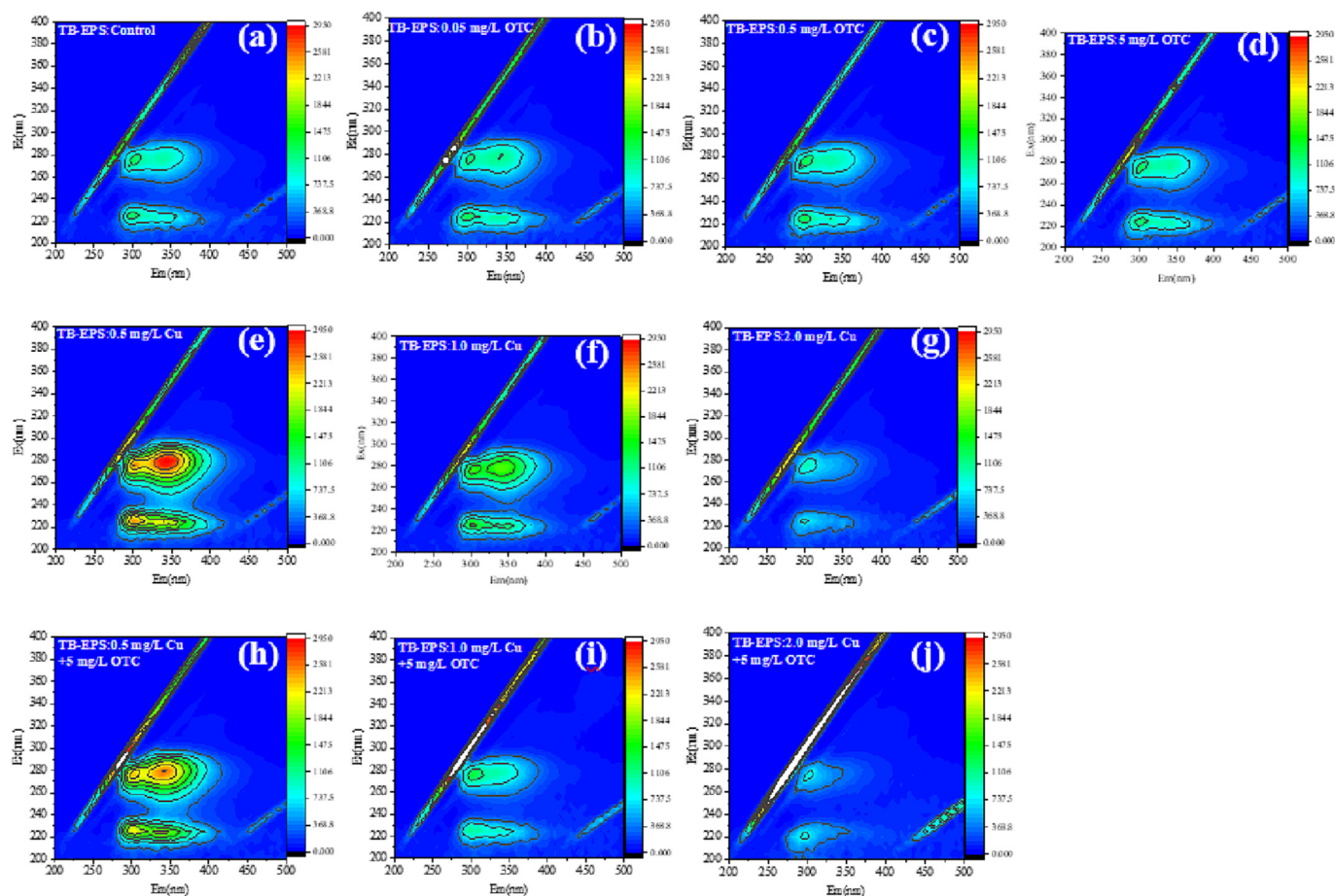
antioxidant enzymes were activated. Moreover, as pollutants increased, microalgae consume ATP to secrete more EPS, thus the ATPase activity also was enhanced.

### 3.6. 3D-EEM of EPS and FTIR analysis

3D-EEM and FTIR are often combined to qualitatively analyze the potential mechanisms by which microorganisms remove heavy metals and antibiotics (Zeeshan et al., 2022). The 3D-EEM spectra of three types of EPS in *Chlorella vulgaris* were extracted. It found that the peaks at 260–280/280–310 (Ex/Em), 260–280/340–360 (Ex/Em), 220–230/280–320 (Ex/Em), 220–230/320–350 (Ex/Em), and 240–250/390–430 (Ex/Em) in the fluorescence spectrum corresponded to tryptophan proteins, aromatic proteins, and humus compounds, respectively (Cui et al., 2021). In S-EPS (Fig. S4A), the fluorescence intensity of each peak enhanced as the stress increased, and the position of the peaks gradually redshifted. This may be that *Chlorella vulgaris* exposed to environmental pressure, the content of proteins and carbohydrates in EPS increased (Fig. S2 and S3) to protect cells from damage (Joshi and Juwarkar, 2009; Wang et al., 2015). Moreover, when environmental stress rose, the growth of microalgae was inhibited, and the increased excretion of microalgae cell due to cells shed and lysed could also be manifested as the fluorescence intensity enhanced in S-EPS (Lapidou and Rittmann, 2002). A study by Oliveira et al. (2023) reported that in high concentration of Zn (25–70 mg/L) with increasing culture time, CODs increased in wastewater were attributed to the increasing secretion of EPS (in the form of carbohydrates), and a strong linear correlation between CODs and EPS was proven. This is a strategy to decline the toxicity on microalgae. In LB-EPS (Fig. S4B), with the concentration of single pollution

increased, the intensity of the peaks gradually enhanced. When combined stress of  $\text{Cu}^{2+}$  and the high concentration of OTC, the fluorescence intensity was weaker. In high concentration, a new peak at 230/300–320 (Ex/Em) appeared, and the main peak in LB-EPS also gradually redshifted. The redshift of the peak indicated that the peak corresponded to the material being more active, which was conducive to the growth of microalgae under environmental pressure (Tang et al., 2020). In TB-EPS (Fig. 6), the fluorescence intensity of each peak gradually declined when the concentration of a single pollutant increased, and further weakened in combined stress. The peak at 260–280/340–360 (Ex/Em) corresponded tryptophan protein gradually disappeared. The fluorescence intensity of TB-EPS weakened, because the content of proteins and carbohydrates increased, and the abundant functional groups (amino and carboxyl groups, etc.) of the proteins may be integrated with  $\text{Cu}^{2+}$  and OTC to produce non-fluorescent chelate (Zhou et al., 2019), particularly the group of  $\text{Cu}^{2+}$  complexed with 5 mg/L of OTC.

By scanning the FTIR spectrum of *Chlorella vulgaris* that ranged from 400 to 4000  $\text{cm}^{-1}$ , the change of functional groups in microalgae was observed (Table 1 and Fig. S5). The multiple spectral bands of these groups have changed in microalgae under stress in experimental group. The peaks at 3305  $\text{cm}^{-1}$ , 2997–3012  $\text{cm}^{-1}$ , 2927  $\text{cm}^{-1}$ , and 1723  $\text{cm}^{-1}$  corresponded to -NH of protein, fatty acids containing -CH<sub>2</sub>, phenolic groups, and carbonyl groups (C=O); and the peaks at 1649–1658  $\text{cm}^{-1}$ , 1539–1546  $\text{cm}^{-1}$ , 1448–1462  $\text{cm}^{-1}$ , 1045–1190  $\text{cm}^{-1}$  were the stretch and vibration of -C=O of protein amide I band, -NH and -C-N groups of protein amide II band, -CH<sub>2</sub>, -CH<sub>3</sub>, and -C-O bonds in lipids and proteins, and C-O-C and C—O bonds of carbohydrates respectively (Cui et al., 2021; Dobrowolski et al., 2017; Liu et al., 2021a; Wei et al., 2017;



**Fig. 6.** Stress of different concentrations of  $\text{Cu}^{2+}$  and OTC on 3D-EEM spectra of TB-EPS of *Chlorella vulgaris* during 12 days culturing in swine wastewater. (a) TB-EPS (control); (b) TB-EPS 0.05 mg/L OTC; (c) TB-EPS 0.5 mg/L OTC; (d) TB-EPS 5 mg/L OTC; (e) TB-EPS 0.5 mg/L  $\text{Cu}^{2+}$ ; (f) TB-EPS 1.0 mg/L  $\text{Cu}^{2+}$ ; (g) TB-EPS 2.0 mg/L  $\text{Cu}^{2+}$ ; (h) TB-EPS 0.5 mg/L  $\text{Cu}^{2+}$  + 5 mg/L OTC; (i) TB-EPS 1.0 mg/L  $\text{Cu}^{2+}$  + 5 mg/L OTC; (j) 2.0 mg/L  $\text{Cu}^{2+}$  + 5 mg/L OTC.

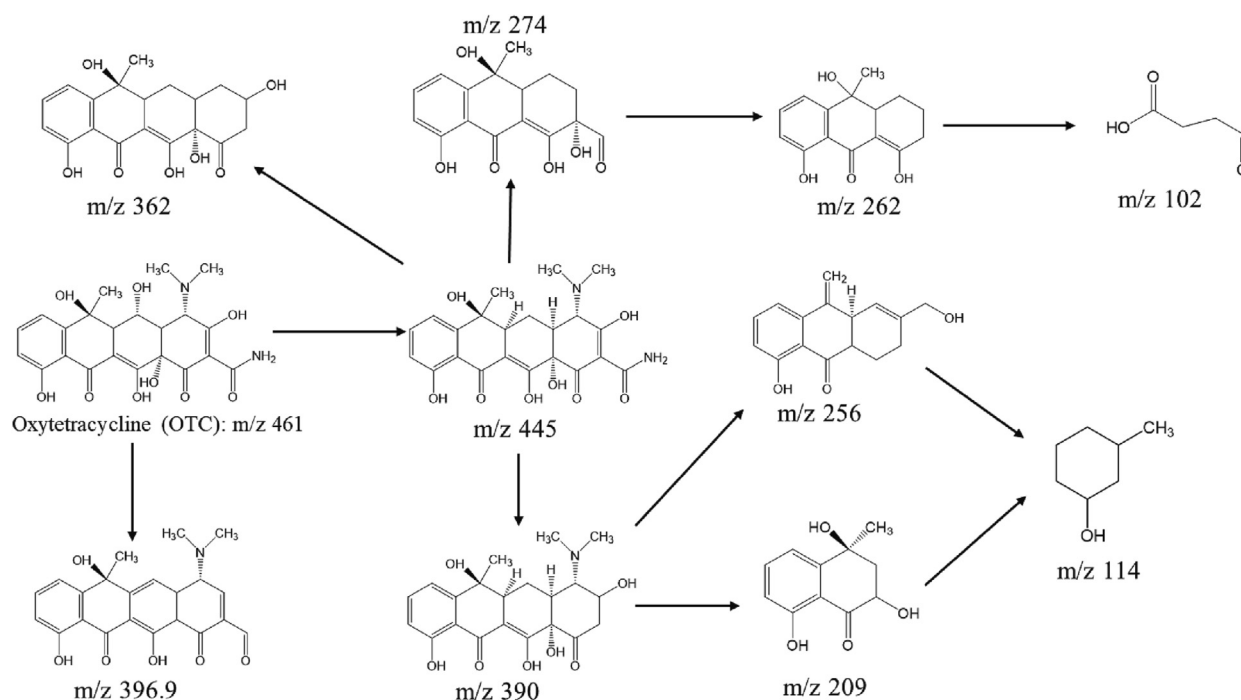
**Table 1**  
Changes of FTIR spectrum of functional groups in *Chlorella vulgaris*.

| Peak      |                  |           |                        | Functional groups                                                           |
|-----------|------------------|-----------|------------------------|-----------------------------------------------------------------------------|
| Control   | Cu <sup>2+</sup> | OTC       | Cu <sup>2+</sup> + OTC |                                                                             |
| 3305      | 3307             | 3303      | 3307                   | -NH of protein                                                              |
| 2297      | 2299–3012        | 3001      | 3001–3012              | -CH <sub>2</sub> of fatty acids                                             |
| 2927      | 2925             | 2927      | 2930                   | phenolic groups                                                             |
| 1723      | 1720             | 1721      | 1720                   | carbonyl group (C=O)                                                        |
| 1651–1656 | 1649–1656        | 1649–1656 | 1649–1658              | -C=O of the protein amide I band                                            |
| 1535–1546 | 1535–1543        | 1535–1242 | 1535–1343              | -NH of protein amide II and -CN groups                                      |
| 1448–1462 | 1446–1460        | 1448–1462 | 1448–1452              | -CH <sub>2</sub> , -CH <sub>3</sub> , and -C-O bonds in lipids and proteins |
| 1045–1190 | 1045–1188        | 1045–1190 | 1045–1182              | C-O-C and C-O bonds of carbohydrates                                        |

Wei et al., 2019). The carboxyl group (-C=O) in the protein made the surface acidic of microalgae and easily expressed a negative charge, which contributed to the adsorption of Cu<sup>2+</sup> and OTC. These proteins and carbohydrates of EPS were also significantly hydrophobic, further promoting the bioabsorption by *Chlorella vulgaris*. The stretch and oscillation of these peaks indicated that Cu<sup>2+</sup> and OTC interacted with functional groups of proteins and carbohydrates in the EPS of microalgae. Liu et al. (2021a) reported that after adsorbing Cu<sup>2+</sup>, the C=O and C=N groups of microalgae at 1644 cm stretched and vibrated, causing the position of the amide I band to change. Moreover, its protein conception changed significantly due to Cu<sup>2+</sup> mainly complexed with carboxy, hydroxyl, and amino groups of TB-EPS. Zeeshan et al. (2022) found that 30 mg/L of antiviral drugs (AVDs) oseltamivir (OT) stimulated the secretion of EPS in *Chlorella vulgaris*. C-S-N and OT were adsorbed and degraded by interacting with C-H groups in carbohydrates, and -NH and -C=O in proteins of EPS.

### 3.7. Degradation pathways of OTC

Oxytetracycline was degraded by *Chlorella vulgaris* and the intermediates were measured by LC-MS (Fig. S6). The intermediates of OTC were shown in Table S4 and the possible degradation pathways were proposed.



**Fig. 7.** The schematic diagram of the degradation pathway of OTC.

Pathway one in the single OTC, OTC (*m/z* 461) generated *m/z* 445 by separating the hydroxide group under the action of the intracellular enzyme of *Chlorella vulgaris*, on the one hand, *m/z* 445 generated *m/z* 362 by demethylation, dehydroxylation, amino separation, and acylamino eliminating; on the other hand, *m/z* 445 generated *m/z* 274 by benzene ring opening, acylamino separating, and hydroxylation, etc., then further dehydroxylation and decarboxylation to generate *m/z* 262, and finally through benzene ring opening, hydroxyl group and methyl decoupling to generate *m/z* 102 (Hu et al., 2019; Li et al., 2022c). Under compound stress, one of the pathways was OTC generated *m/z* 396.9 by separating three hydroxyl groups and one amino group. Another pathway was the hydroxyl group of OTC separated to form *m/z* 445, after the acylamino and hydroxyl group detached to form *m/z* 390, which was degraded through two pathways in turn. One was to generate *m/z* 256 by carbon ring decarboxylation and splitting decomposition, dehydrogenation and dehydration, and finally, benzene ring cracking to generate *m/z* 114; the second was hydroxyl isolated, methylation, dehydrogenation, -N(CH<sub>3</sub>)<sub>2</sub> broken away, and benzene ring directly cleavage to generate *m/z* 209, and finally produce *m/z* 114 (Lian et al., 2021; Liu et al., 2022). The schematic diagram of the degradation pathway of OTC was shown in Fig. 7. Finally, there were five possible complexed forms that Cu<sup>2+</sup> complexed with OTC in swine wastewater shown in Table S5 (Zhang et al., 2021b; Zhang et al., 2012).

### 4. Conclusions

Dynamic hormesis of either OTC concentration or Cu<sup>2+</sup> one on *Chlorella vulgaris* growth were confirmed separately. At appropriate concentration and culture time, OTC promoted the NH<sub>3</sub>-N removal, biomass growth, and lipids content of *Chlorella vulgaris*. At the low concentration of Cu<sup>2+</sup> (≤1.0 mg/L), the content of protein and SOD increased, however, 2.0 mg/L of Cu<sup>2+</sup> could cause severe inhibition of *Chlorella vulgaris* growth and the accumulation of lipids. Interestingly, the content of lipids, GHS, and activity of ATPase began to increase, and H<sub>2</sub>O<sub>2</sub> content decreased under combined stress of Cu<sup>2+</sup> and OTC, and the presence of OTC helped to alleviate the toxicity caused by Cu<sup>2+</sup>. Nevertheless, combined stress of Cu<sup>2+</sup> and OTC expressed insignificant for promoting the removal of NH<sub>3</sub>-N. Cu<sup>2+</sup> and OTC first complexed in swine wastewater, which reduced the toxic effect of Cu<sup>2+</sup> on *Chlorella vulgaris*. Cu<sup>2+</sup> and OTC bonded with the

group of proteins in TB-EPS to produce the chelate of the non-fluorescence characteristics, which promoted mutually adsorbed by *Chlorella vulgaris*.

### CRedit authorship contribution statement

**Yun Luo:** Conceptualization, Methodology, Investigation, Writing-Original draft preparation, Writing-Reviewing and Editing. **Xiang Li:** Conceptualization, Methodology, Investigation, Writing-original draft, Writing-Reviewing and Editing. **Yan Lin:** Conceptualization, Methodology, Data curation, Writing-Original draft preparation, Writing-Reviewing and Editing. **Shaohua Wu:** Visualization, Investigation, Validation, Writing-Original draft preparation. **Jay J. Cheng:** Methodology, Writing-Original draft preparation. **Chunping Yang:** Conceptualization, Supervision, Project administration, Funding acquisition, Writing-Original draft preparation, Writing-Reviewing and Editing.

### Data availability

Data will be made available on request.

### Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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### Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.scitotenv.2023.165120>.

### References

- Agathokleous, E., Calabrese, E.J., 2022. Hormesis: a general biological principle. *Chem. Res. Toxicol.* 35 (4), 547–549.
- Agathokleous, E., Barceló, D., Rinklebe, J., Sonne, C., Calabrese, E.J., Koike, T., 2022. Hormesis induced by silver iodide, hydrocarbons, microplastics, pesticides, and pharmaceuticals: implications for agroforestry ecosystems health. *Sci. Total Environ.* 820, 153116.
- Bahr, M., Díaz, I., Domínguez, A., González Sánchez, A., Muñoz, R., 2014. Microalgal-biotechnology as a platform for an integral biogas upgrading and nutrient removal from anaerobic effluents. *Environ. Sci. Technol.* 48 (1), 573–581.
- Bechaieb, R., Ben Akacha, A., Gérard, H., 2016. Quantum chemistry insight into mg-substitution in chlorophyll by toxic heavy metals: Cd, Hg and Pb. *Chem. Phys. Lett.* 663, 27–32.
- Ben, W., Pan, X., Qiang, Z., 2013. Occurrence and partition of antibiotics in the liquid and solid phases of swine wastewater from concentrated animal feeding operations in Shandong province, China. *Environ. Sci.: Processes Impacts* 15 (4), 870–875.
- Cestonaro do Amaral, A., Kunz, A., Radis Steinmetz, R.L., Justi, K.C., 2014. Zinc and copper distribution in swine wastewater treated by anaerobic digestion. *J. Environ. Manag.* 141, 132–137.
- Chen, K., Wu, X., Zou, Z., Dong, Y., Zhang, S., Li, X., Gouda, M., Chu, B., Li, C.M., Li, X., He, Y., 2022. Assess heavy metals-induced oxidative stress of microalgae by electro-raman combined technique. *Anal. Chim. Acta* 1208, 339791.
- Cheng, J., Ye, Q., Li, K., Liu, J., Zhou, J., 2018. Removing ethinylestradiol from wastewater by microalgae mutant *Chlorella PY-ZUI* with CO<sub>2</sub> fixation. *Bioresour. Technol.* 249, 284–289.
- Cheng, D.L., Ngo, H.H., Guo, W.S., Chang, S.W., Nguyen, D.D., Kumar, S.M., 2019. Microalgae biomass from swine wastewater and its conversion to bioenergy. *Bioresour. Technol.* 275, 109–122.
- Christenson, L., Sims, R., 2011. Production and harvesting of microalgae for wastewater treatment, biofuels, and bioproducts. *Biotechnol. Adv.* 29 (6), 686–702.
- Chu, Y., Zhang, C., Chen, X., Li, X., Ren, N., Ho, S.-H., 2023. Multistage defense response of microalgae exposed to pharmaceuticals in wastewater. *Chin. Chem. Lett.* 34 (4), 107727.
- Cui, L., Fan, L., Li, Z., Wang, J., Chen, R., Zhang, Y., Cheng, J., Wu, X., Li, J., Yin, H., Zeng, W., Shen, L., 2021. Characterization of extracellular polymeric substances from *Synechocystis* sp. PCC6803 under Cd (II), Pb (II) and Cr (VI) stress. *J. Environ. Chem. Eng.* 9 (4), 105347.
- D'Abzac, P., Bordes, F., Van Hullebusch, E., Lens, P.N.L., Guibaud, G., 2010. Extraction of extracellular polymeric substances (EPS) from anaerobic granular sludges: comparison of chemical and physical extraction protocols. *Appl. Microbiol. Biotechnol.* 85 (5), 1589–1599.
- Daneshvar, E., Antikainen, L., Koutra, E., Komaros, M., Bhatnagar, A., 2018. Investigation on the feasibility of *Chlorella vulgaris* cultivation in a mixture of pulp and aquaculture effluents: treatment of wastewater and lipid extraction. *Bioresour. Technol.* 255, 104–110.
- Danouche, M., El Ghachtouli, N., El Baouchi, A., El Arroussi, H., 2020. Heavy metals phytoremediation using tolerant green microalgae: enzymatic and non-enzymatic antioxidant systems for the management of oxidative stress. *J. Environ. Chem. Eng.* 8 (5), 104460.
- Demidchik, V., Straltsova, D., Medvedev, S.S., Pozhvanov, G.A., Sokolik, A., Yurin, V., 2014. Stress-induced electrolyte leakage: the role of K<sup>+</sup>-permeable channels and involvement in programmed cell death and metabolic adjustment. *J. Exp. Bot.* 65 (5), 1259–1270.
- Dobrowolski, R., Szcześ, A., Czemierska, M., Jarosz-Wikolazka, A., 2017. Studies of cadmium (II), lead(II), nickel(II), cobalt(II) and chromium(VI) sorption on extracellular polymeric substances produced by *rhodococcus opacus* and *rhodococcus rhodochrous*. *Bioresour. Technol.* 225, 113–120.
- Edreva, A., 2005. Generation and scavenging of reactive oxygen species in chloroplasts: a submolecular approach. *Agric. Ecosyst. Environ.* 106 (2), 119–133.
- Erofeeva, E.A., 2022. Environmental hormesis: from cell to ecosystem. *Curr. Opin. Environ. Sci. Health* 29, 100378.
- Hamed, S.M., Selim, S., Klöck, G., Abdelgawad, H., 2017. Sensitivity of two green microalgae to copper stress: growth, oxidative and antioxidants analyses. *Ecotox. Environ. Safe.* 144, 19–25.
- Hu, H., Zhou, Q., Li, X., Lou, W., Du, C., Teng, Q., Zhang, D., Liu, H., Zhong, Y., Yang, C., 2019. Phytoremediation of anaerobically digested swine wastewater contaminated by oxytetracycline via *Lemna aequinoctialis*: nutrient removal, growth characteristics and degradation pathways. *Bioresour. Technol.* 291, 121853.
- Hu, H., Li, X., Wu, S., Yang, C., 2020. Sustainable livestock wastewater treatment via phytoremediation: current status and future perspectives. *Bioresour. Technol.* 315, 123809.
- Hu, H., Li, X., Wu, S., Lou, W., Yang, C., 2021. Effects of long-term exposure to oxytetracycline on phytoremediation of swine wastewater via duckweed systems. *J. Hazard. Mater.* 414, 125508.
- Joshi, P.M., Juwarkar, A.A., 2009. In vivo studies to elucidate the role of extracellular polymeric substances from azotobacter in immobilization of heavy metals. *Environ. Sci. Technol.* 43 (15), 5884–5889.
- Khan, S.J., Roser, D.J., Davies, C.M., Peters, G.M., Stuetz, R.M., Tucker, R., Ashbolt, N.J., 2008. Chemical contaminants in feedlot wastes: concentrations, effects and attenuation. *Environ. Int.* 34 (6), 839–859.
- Kumaran, M., Palanisamy, K.M., Bhuyar, P., Maniam, G.P., Rahim, M.H.A., Govindan, N., 2023. Agriculture of microalgae *Chlorella vulgaris* for polyunsaturated fatty acids (PUFAs) production employing palm oil mill effluents (POME) for future food, wastewater, and energy nexus. *Energy Nexus* 9, 100169.
- Laspidou, C.S., Rittmann, B.E., 2002. A unified theory for extracellular polymeric substances, soluble microbial products, and active and inert biomass. *Water Res.* 36 (11), 2711–2720.
- Li, X., Yang, W.L., He, H., Wu, S., Zhou, Q., Yang, C., Zeng, G., Luo, L., Lou, W., 2018. Responses of microalgae *Coelastrella* sp. to stress of cupric ions in treatment of anaerobically digested swine wastewater. *Bioresour. Technol.* 251, 274–279.
- Li, S., Chu, R., Hu, D., Yin, Z., Mo, F., Hu, T., Liu, C., Zhu, L., 2020a. Combined effects of 17β-estradiol and copper on growth, biochemical characteristics and pollutant removals of freshwater microalgae *Scenedesmus dimorphus*. *Sci. Total Environ.* 730, 138597.
- Li, X., Yang, C., Zeng, G., Wu, S., Lin, Y., Zhou, Q., Lou, W., Du, C., Nie, L., Zhong, Y., 2020b. Nutrient removal from swine wastewater with growing microalgae at various zinc concentrations. *Algal Res.* 46, 101804.
- Li, X., Gu, D., You, J., Qiao, T., Yu, X., 2022a. Gamma-aminobutyric acid coupled with copper ion stress stimulates lipid production of green microalga *Monoraphidium* sp. QLY-1 through multiple mechanisms. *Bioresour. Technol.* 352, 127091.
- Li, X., Lu, Y., Li, N., Wang, Y., Yu, R., Zhu, G., Zeng, R.J., 2022b. Mixotrophic cultivation of microalgae using biogas as the substrate. *Environ. Sci. Technol.* 56 (6), 3669–3677.
- Li, X., Yang, C., Lin, Y., Hu, T., Zeng, G., 2022c. Effects of oxytetracycline and zinc ion on nutrient removal and biomass production via microalgal culturing in anaerobic digester effluent. *Bioresour. Technol.* 346, 126667.
- Li, Y., Qiu, Z., Cui, H., Wang, L., Zhang, L., 2022d. Simultaneous promotion of microalgal CO<sub>2</sub> assimilation, biomass accumulation, lipid production, and wastewater nutrient removal by adding 5-aminolevulinic acid. *ACS Sustain. Chem. Eng.* 10 (45), 14715–14723.
- Li, C., Lin, Y., Li, X., Cheng, J.J., Yang, C., 2023. Cupric ions inducing dynamic hormesis in duckweed systems for swine wastewater treatment: quantification, modelling and mechanisms. *Sci. Total Environ.* 866, 161411.
- Lian, S., Shi, X., Lu, M., Zhang, M., Dong, X., Li, X., Feng, Q., Guo, R., 2021. Accelerated adsorption of tetracyclines and microbes with Fe(OH)<sub>3</sub> modified oyster shell: its application on biotransformation of oxytetracycline in anaerobic enrichment culture. *Chem. Eng. J.* 425, 130499.
- Lin, Y., Wu, X., Han, Y., Yang, C., Ma, Y., Du, C., Teng, Q., Liu, H., Zhong, Y., 2019. Spatial separation of photogenerated carriers and enhanced photocatalytic performance on Ag<sub>3</sub>PO<sub>4</sub> catalysts via coupling with PPy and MWCNTs. *Appl. Catal. B-Environ.* 258, 117969.



- Liu, L., Lin, X., Luo, L., Yang, J., Luo, J., Liao, X., Cheng, H., 2021a. Biosorption of copper ions through microalgae from piggy digestate: optimization, kinetic, isotherm and mechanism. *J. Clean. Prod.* 319, 128724.
- Liu, T., Wang, Y., Li, J., Yu, Q., Wang, X., Gao, D., Wang, F., Cai, S., Zeng, Y., 2021b. Effects from Fe, P, Ca, Mg, Zn and Cu in steel slag on growth and metabolite accumulation of microalgae: A review. *Appl. Sci.* 11 (14), 6589.
- Liu, X., Pei, Y., Cao, M., Yang, H., Li, Y., 2022. Highly dispersed copper single-atom catalysts activated peroxymonosulfate for oxytetracycline removal from water: mechanism and degradation pathway. *Chem. Eng. J.* 450, 138194.
- Lu, L., Wu, Y., Ding, H., Zhang, W., 2015. The combined and second exposure effect of copper (II) and chlortetracycline on fresh water algae, *Chlorella pyrenoidosa* and *Microcystis aeruginosa*. *Environ. Toxicol. Pharmacol.* 40 (1), 140–148.
- Luo, L., He, H., Yang, C., Wen, S., Zeng, G., Wu, M., Zhou, Z., Lou, W., 2016. Nutrient removal and lipid production by *Coelastrella* sp. in anaerobically and aerobically treated swine wastewater. *Bioresour. Technol.* 216, 135–141.
- Martínez-Macias, M.d.R., Correa-Murrieta, M.A., Villegas-Peralta, Y., Dévora-Isordia, G.E., Álvarez-Sánchez, J., Saldivar-Cabrera, J., Sánchez-Duarte, R.G., 2019. Uptake of copper from acid mine drainage by the microalgae *Nannochloropsis oculata*. *Environ. Sci. Pollut. Res.* 26 (7), 6311–6318.
- Miazek, K., Iwanek, W., Remacle, C., Richel, A., Goffin, D., 2015. Effect of metals, metalloids and metallic nanoparticles on microalgae growth and industrial product biosynthesis: A review. *Int. J. Mol. Sci.* 16, 23929–23969.
- Michelon, W., Matthiensen, A., Viancelli, A., Fongaro, G., Gressler, V., Soares, H.M., 2022. Removal of veterinary antibiotics in swine wastewater using microalgae-based process. *Environ. Res.* 207, 112192.
- Mishra, B., Chandra, M., Pant, D., 2021. Genome-mining for stress-responsive genes, profiling of antioxidants and radical scavenging metabolism in hyperaccumulator medicinal and aromatic plants. *Ind. Crop. Prod.* 173, 114107.
- Nicodemus, T.J., DiRusso, C.C., Wilson, M., Black, P.N., 2020. Reactive oxygen species (ROS) mediated degradation of organophosphate pesticides by the green microalgae *Coccomyxa subellipsoidea*. *Bioresour. Technol.* 216, 100461.
- Noctor, G., Arisi, A.-C.M., Jouanin, L., Foyer, C.H., 1998. Manipulation of glutathione and amino acid biosynthesis in the *Chloroplast*. *Plant Physiol.* 118 (2), 471–482.
- Okamoto, O.K., Pinto, E., Latorre, L.R., Bechara, E.J.H., Colepicolo, P., 2001. Antioxidant modulation in response to metal-induced oxidative stress in algal *chloroplasts*. *Arch. Environ. Contam. Toxicol.* 40 (1), 18–24.
- Oliveira, A.P.d.S., Assemany, P., Covell, L., Tavares, G.P., Calijuri, M.L., 2023. Microalgae-based wastewater treatment for micropollutant removal in swine effluent: high-rate algal ponds performance under different zinc concentrations. *Algal Res.* 69, 102930.
- Perales-Vela, H.V., Peña-Castro, J.M., Cañizares-Villanueva, R.O., 2006. Heavy metal detoxification in eukaryotic microalgae. *Chemosphere* 64 (1), 1–10.
- Qin, L., Feng, P., Zhu, S., Xu, Z., Wang, Z., 2022. A novel partial complete nitrification coupled microalgae assimilation system for resource utilization of high ammonia nitrogen wastewater. *J. Environ. Chem. Eng.* 10 (6), 108584.
- Sabatini, S.E., Juárez, Á.B., Eppis, M.R., Bianchi, L., Luquet, C.M., de Molina, Ríos, M.d.C., 2009. Oxidative stress and antioxidant defenses in two green microalgae exposed to copper. *Ecotox. Environ. Safe.* 72 (4), 1200–1206.
- Serejo, M.L., Posadas, E., Boncz, M.A., Blanco, S., García-Encina, P., Muñoz, R., 2015. Influence of biogas flow rate on biomass composition during the optimization of biogas upgrading in microalgal-bacterial processes. *Environ. Sci. Technol.* 49 (5), 3228–3236.
- Shen, Q.-H., Gong, Y.-P., Fang, W.-Z., Bi, Z.-C., Cheng, L.-H., Xu, X.-H., Chen, H.-L., 2015. Saline wastewater treatment by *Chlorella vulgaris* with simultaneous algal lipid accumulation triggered by nitrate deficiency. *Bioresour. Technol.* 193, 68–75.
- Sheng, G.-P., Yu, H.-Q., Li, X.-Y., 2010. Extracellular polymeric substances (EPS) of microbial aggregates in biological wastewater treatment systems: a review. *Biotechnol. Adv.* 28 (6), 882–894.
- Siedlewiez, G., Żak, A., Sharma, L., Kosakowska, A., Pazdro, K., 2020. Effects of oxytetracycline on growth and chlorophyll a fluorescence in green algae (*Chlorella vulgaris*), diatom (*Phaeodactylum tricornutum*) and cyanobacteria (*Microcystis aeruginosa* and *Nodularia spumigena*). *Oceanologia* 62 (2), 214–225.
- Suzuki, K., Waki, M., Yasuda, T., Fukumoto, Y., Kuroda, K., Sakai, T., Suzuki, N., Suzuki, R., Matsuba, K., 2010. Distribution of phosphorus, copper and zinc in activated sludge treatment process of swine wastewater. *Bioresour. Technol.* 101 (23), 9399–9404.
- Tang, W., Li, X., Liu, H., Wu, S., Zhou, Q., Du, C., Teng, Q., Zhong, Y., Yang, C., 2020. Sequential vertical flow trickling filter and horizontal flow multi-soil-layering reactor for treatment of decentralized domestic wastewater with sodium dodecyl benzene sulfonate. *Bioresour. Technol.* 300, 122634.
- Upadhyay, A.K., Mandotra, S.K., Kumar, N., Singh, N.K., Singh, L., Rai, U.N., 2016. Augmentation of arsenic enhances lipid yield and defense responses in alga *Nannochloropsis* sp. *Bioresour. Technol.* 221, 430–437.
- Verma, R., Suthar, S., Chand, N., Mutiyar, P.K., 2022. Phycoremediation of milk processing wastewater and lipid-rich biomass production using *Chlorella vulgaris* under continuous batch system. *Sci. Total Environ.* 833, 155110.
- Wang, Y., Qin, J., Zhou, S., Lin, X., Ye, L., Song, C., Yan, Y., 2015. Identification of the function of extracellular polymeric substances (EPS) in denitrifying phosphorus removal sludge in the presence of copper ion. *Water Res.* 73, 252–264.
- Wang, J., Ben, W., Yang, M., Zhang, Y., Qiang, Z., 2016. Dissemination of veterinary antibiotics and corresponding resistance genes from a concentrated swine feedlot along the waste treatment paths. *Environ. Int.* 92–93, 317–323.
- Wang, X., Dou, X., Wu, J., Meng, F., 2021. Attenuation pathways of erythromycin and biochemical responses related to algal growth and lipid synthesis in a microalga-effluent system. *Environ. Res.* 195, 110873.
- Wei, L., Li, Y., Noguera, D.R., Zhao, N., Song, Y., Ding, J., Zhao, Q., Cui, F., 2017. Adsorption of  $\text{Cu}^{2+}$  and  $\text{Zn}^{2+}$  by extracellular polymeric substances (EPS) in different sludges: effect of EPS fractional polarity on binding mechanism. *J. Hazard. Mater.* 321, 473–483.
- Wei, L., Li, J., Xue, M., Wang, S., Li, Q., Qin, K., Jiang, J., Ding, J., Zhao, Q., 2019. Adsorption behaviors of  $\text{Cu}^{2+}$ ,  $\text{Zn}^{2+}$  and  $\text{Cd}^{2+}$  onto proteins, humic acid, and polysaccharides extracted from sludge EPS: sorption properties and mechanisms. *Bioresour. Technol.* 291, 121868.
- Wu, C., He, C., 2019. Interaction effects of oxytetracycline and copper at different ratios on marine microalgae *Isochrysis galbana*. *Chemosphere* 225, 775–784.
- Wu, S., Zhang, J., Xia, A., Huang, Y., Zhu, X., Zhu, X., Liao, Q., 2022. Microalgae cultivation for antibiotic oxytetracycline wastewater treatment. *Environ. Res.* 214, 113850.
- Wu, X., Lin, Y., Wang, Y., Wu, S., Li, X., Yang, C., 2022b. Enhanced removal of hydrophobic short-chain n-alkanes from gas streams in biotrickling filters in presence of surfactant. *Environ. Sci. Technol.* 56 (14), 10349–10360.
- Xia, A., Murphy, J.D., 2016. Microalgal cultivation in treating liquid digestate from biogas systems. *Trends Biotechnol.* 34 (4), 264–275.
- Xiao, Y., Yang, C., Cheng, J.J., 2022. Effects of sulfamethazine and cupric ion on treatment of anaerobically digested swine wastewater with growing duckweed. *Int. J. Environ. Res. Public Health* 19 (4), 1949.
- Xiao, X., Li, W., Jin, M., Zhang, L., Qin, L., Geng, W., 2023. Responses and tolerance mechanisms of microalgae to heavy metal stress: A review. *Mar. Environ. Res.* 183, 105805.
- Xie, P., Ho, S.-H., Peng, J., Xu, X.-J., Chen, C., Zhang, Z.-F., Lee, D.-J., Ren, N.-Q., 2019. Dual purpose microalgae-based biorefinery for treating pharmaceuticals and personal care products (PPCPs) residues and biodiesel production. *Sci. Total Environ.* 688, 253–261.
- Xue, J., Balamurugan, S., Li, D.-W., Liu, Y.-H., Zeng, H., Wang, L., Yang, W.-D., Liu, J.-S., Li, H.-Y., 2017. Glucose-6-phosphate dehydrogenase as a target for highly efficient fatty acid biosynthesis in microalgae by enhancing NADPH supply. *Metab. Eng.* 41, 212–221.
- Yang, H., Zhao, Z., Liu, Y., Fu, L., Zhou, D., 2023. The p-hydroxybenzoic acid enhanced lipid accumulation of *Chlorella* under antibiotic stress. *Resour. Conserv. Recycl.* 190, 106758.
- You, X., Li, H., Pan, B., You, M., Sun, W., 2022. Interactions between antibiotics and heavy metals determine their combined toxicity to *Synechocystis* sp. *J. Hazard. Mater.* 424, 127707.
- Zeeshan, Q.M., Qiu, S., Gu, J., Abbew, A.-W., Wu, Z., Chen, Z., Xu, S., Ge, S., 2022. Unravelling multiple removal pathways of oseltamivir in wastewater by microalgae through experimentation and computation. *J. Hazard. Mater.* 427, 128139.
- Zhang, Y., Cai, X., Lang, X., Qiao, X., Li, X., Chen, J., 2012. Insights into aquatic toxicities of the antibiotics oxytetracycline and ciprofloxacin in the presence of metal: complexation versus mixture. *Environ. Pollut.* 166, 48–56.
- Zhang, H., Xu, Z., Huo, Y., Guo, K., Wang, Y., He, G., Sun, H., Li, M., Li, X., Xu, N., Sun, G., 2020. Overexpression of Trx CDSP32 gene promotes chlorophyll synthesis and photosynthetic electron transfer and alleviates cadmium-induced photoinhibition of PSII and PSI in tobacco leaves. *J. Hazard. Mater.* 398, 122899.
- Zhang, C., Li, R., Feng, Y., Zhang, L., Zhao, C., Ji, C., Cui, H., 2021a. Glycinebetaine promotes photosynthesis, biomass accumulation, and lipid production in *nannochloropsis gaditana* under nitrogen deprivation. *ACS Sustain. Chem. Eng.* 9 (51), 17232–17241.
- Zhang, Q.-Q., Qian, H., Li, P.-Y., Zhao, J.-Q., Sun, Y.-Q., Jin, R.-C., 2021b. Insight into the evolution of microbial community and antibiotic resistance genes in anammox process induced by copper after recovery from oxytetracycline stress. *Bioresour. Technol.* 330, 124945.
- Zhao, Y., Tan, Y., Guo, Y., Gu, X., Wang, X., Zhang, Y., 2013. Interactions of tetracycline with Cd (II), Cu (II) and Pb (II) and their cosorption behavior in soils. *Environ. Pollut.* 180, 206–213.
- Zhou, Q., Lin, Y., Li, X., Yang, C., Han, Z., Zeng, G., Lu, L., He, S., 2018. Effect of zinc ions on nutrient removal and growth of *Lemna aequinoctialis* from anaerobically digested swine wastewater. *Bioresour. Technol.* 249, 457–463.
- Zhou, Q., Li, X., Lin, Y., Yang, C., Tang, W., Wu, S., Li, D., Lou, W., 2019. Effects of copper ions on removal of nutrients from swine wastewater and on release of dissolved organic matter in duckweed systems. *Water Res.* 158, 171–181.