- Waste valorization: Transforming the fishbone biowaste into
- 2 biochar as an efficient persulfate catalyst for degradation of
- 3 organic pollutant
- 4 Xiaoya Ren a,b, Jingjing Wang a,b, Jiangfang Yu a,b, Biao Song a,b, Haopeng Feng a,b, Maocai
- 5 Shen a,b, Hao Zhang a,b, Jiajing Zou a,b, Guangming Zeng a,b, Lin Tang a,b, Jiajia Wang a,b,
- 6 ^a College of Environmental Science and Engineering, Hunan University, Changsha,
- 7 410082, China;

- 8 b Key Laboratory of Environmental Biology and Pollution Control, Hunan
- 9 University, Ministry of Education, Changsha 410082, China

Kccelyrey

E-mail: zgming@hnu.edu.cn (Guangming Zeng), tanglin@hnu.edu.cn (Lin Tang), wangjiajia07@hnu.edu.cn (Jiajia Wang)

^{*} Corresponding Authors

Abstract

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Fishbone is a major by-product of fishing processing and how to deal with the increasingly generated fishbone has been a big challenge. In an effort to convert the low-value waste into valuable material, this research explores the valorization of fishbone biowaste, which can be transformed into biochar as a persulfate activator for phenol removal. The fishbone derived biochar (FBBC) was prepared by one-step pyrolysis without additional template. The catalytic efficiency of FBBC increases with pyrolysis temperature, with FBBC-800 (pyrolyzed at 800) exhibiting the best performance of 100% phenol (20 mg/L) removal with hin at the catalyst dosage of 0.1 g/L, which is comparable to other cently reported carbon-based catalysts but with the advantage of easy programon, low cost and wastes valorization. The high efficacy of FBBC-300 is related to its large surface area efective structure. In the FBBC-800/ $(758.44 \text{ m}^2/\text{g})$, carbonyl grou peroxydisulfate /phenol sys em, both radical and non-radical pathways are involved, adi al is more important in radical pathway while singlet among which hy oxygen dominates in non-radical pathway. Electron transfer plays a key role in this process through electron capture experiment and electrochemical characterization. This study proposes a new strategy for the valorization of fishbone, and provides the guidance for the structure design of carbon-based catalyst for persulfate activation.

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- **Keywords**: Waste valorization, Calcium rich biowaste, Fishbone biochar, Persulfate
- 32 activation

1. Introduction

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The global fish production has been increasing over recent years, from 20 35 36 million tons in 1950 up to 179 million tons in 2018, and is projected to reach 204 million tons in 2030 (FAO, 2020). This is accompanied with a large amount of 37 38 fishery wastes, with a estimation of over 20 million tons per year (Nawaz et al., 2020). Fishbone is the main by-product from fish processing, accounting for 39 8%-10% of total fish biomass (Yin et al., 2015). In many cases, fishbone is 40 discarded directly or used for low value fish meal production, where the directly of used for low value fish meal production, where the discarded directly or used for low value fish meal production, where the discarded directly or used for low value fish meal production, where the discarded directly or used for low value fish meal production. 41 is a waste of resources (Marti-Quijal et al., 2020). Some efforts have be on extraction of 42 substance (e.g. hydroxyapatite, gelatin and bioactive alcium, etc) from fishbone for 43 human consumption (Akag ünd üz et al., 2014; 30 linguiza et al., 2012; Zhang et al., 44 2016). However, it is still challenging for large-scale application and the food safety 45 al., 2020). Pyrolysis is a good way to issues are seriously concerned 46 shbone as it is convenient and simple, and the resultant achieve the valorization of 47 de eloped for various applications (Kim et al., 2020). biochar product 48 Fishbone is a natural composite of organic (21%-57%) and inorganic substance 49 (31%-73%) (Toppe et al., 2007). The organic portions such as collagen, proteins 50 and fats serve as carbon precursors, and nanohydroxyapatite, the major inorganic 51 52 mineral present in bone act as self-template. Such unique characteristics make it a perfect material for biochar preparation. Although previous studies have explored 53 54 the application of fishbone in the catalyst reaction, either it is used as a catalyst support (Chakraborty and RoyChowdhury, 2013) or some chemical agents (e.g. 55

KOH and FeCl₃) are used during the catalyst preparation process (Wang et al. 2015; Niu et al. 2019a). The inherent catalytic ability was ignored and the complicated production process would also increase the costs. This study makes full use of the self-template in fishbone to prepare porous carbon catalyst, and the preparation process is simple and environmental friendly. In addition, currently, fishbone derived biochar (FBBC) has been applied in electro-catalysis, supercapacitor and pollutant adsorption (Medell ń-Castillo et al., 2020; Niu et al., 2019a; Wang et al., 2015), which proves the possible adsorption and catalytic ability. However, utilizing FBBC as a catalyst for pollutant degradation has rarely been reported. As a good electron donor and electron carrier, FBBC hight be exploited in the persulfate-induced advanced oxidation processes (S-AOPs).

The PS-AOPs is a kind of water treatment technology and has attracted more and more interest in recent year one is as easy operation and high mineralization efficiency. Peroxydisulfate (PDS) is a stable oxidant with low oxidative potential for pollutant degrecation (Waclawek et al., 2017). However, the peroxide O-O bond in PDS can be cleaved with the assistance of catalysts or external energy, along with the generation of highly reactive species including hydroxyl radicals (OH) and sulfate radicals (SO₄ ·), which enables the effective degradation of refractory organics (Duan et al., 2015). Various methods have been used for PDS activation, such as UV irradiation, thermal, ultrasound, transition metals and carbonaceous materials (Wang and Wang, 2018). Among these methods, biochar exhibits unique benefits of low cost, rich feedstock sources and simple preparation process (Ye et al.,

2019). However, to overcome the problem of unsatisfactory catalytic ability in pristine biochar, heteroatom or metal doping have been applied to improve the reactivity of biochar (Fang et al., 2015; Wang et al., 2019a), but this comes out at the cost of excessive synthesis cost or metal leaching. Consequently, attention has been directed toward the structure and chemical constitution of biomass for biochar production. For example, Chen et al. (2019) employed Fe/N carbocatalyst Enteromorpha biomass preparing for and used peroxymonosulfate activation. The intrinsic N derived from E *orpha* provide the active site for paracetamol degradation. Similarly, Ma 018) synthesized N/S co-doped porous carbon from human hair, while exhibited better degradation efficiency of bisphenol A than some metal-ba ed atarysts in PDS system. However, KOH as the activator was used in those studies to form porous structure, which is corrosive to the equipment. Bio abundant elements such as K, Na and Ca is a good choice of biocha raw material, which provide the possibility of in-situ s process. activation during Inspired by above results, we believe that FBBC would exhibit excellent

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activity for PDS activation. The porous structure derived from nanohydroxyapatite self-template is favorable for the catalytic reaction (Alhokbany et al., 2019). On the one hand, the well-developed porous structure exposes more active site for catalysis. On the other hand, the intensified adsorption of organic pollutants around the active site facilitate the subsequent catalytic degradation. In addition, electron transfer is an important process in the catalytic reaction (Ren et al., 2019; Yang et al., 2018).

Wang et al. (2015) have studied the electrocatalytic performance of fishbone carbon in the oxygen reduction reaction, and it showed high electron transfer capacity.

Based on the peculiar component of fishbone and the concept of waste valorization, we choose fishbone as a raw material to prepare biochar. The FBBC was applied to activate PDS for degradation of a model pollutant (phenol), where the impact of biochar pyrolysis temperature was studied. The possible catalytic site responsible for the activation reaction was explored. Radical quenching experiments, electron spin resonance and electrochemical characterization w conducted to reveal the activation mechanism. The removal efficiency I in real samples were also investigated. This research proposes a new pute for fishbone valorization, with both the advantage of reducing biowast dep sal cost and creating effective catalyst for pollutant removal. It is expected that this study would shed light on the biomass selection and structura on of biochar to get excellent catalytic efficiency.

2. Materials and methods

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In this section firstly, the specific information about the chemicals and materials used in this study is provided. Secondly, the preparation process of FBBC catalyst is described. Then the techniques used to characterize the structural properties of FBBC are summarized. At last, the detailed reaction procedure and reaction parameters are presented.

2.1. Chemicals and materials

The abyssal fishbone was obtained from a fish processing factory in Weihai,

China. All the chemicals used in this study were used at analytical grade (details can be seen in Text S1). Ultrapure water (18.25 M Ω /cm) was used throughout the experiment.

2.2. Preparation of biochar

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FBBC was prepared as described previously with some modification (Wang et al. 2015). Fishbone was washed with deionized water for several times and dried in the oven for 24 h. Then the dried sample was smashed into powder, screened by 100 mesh sieves and pyrolyzed in a quartz tube furnace at d fee temperatures (400 $\,^{\circ}$ C, 600 $\,^{\circ}$ C and 800 $\,^{\circ}$ C) for 2 h under nitrogen environ itrogen flow rate of 100 mL/min, heating rate of 5 °C/min). After calling to room temperature, the obtained product was washed with 2M H(1) h to remove the calcium compounds. Subsequently, the product was kneed by ultrapure water until the pH of filtrate present neutrality and ℃ for 24 h. The final sample was designated as FBBC-400 FBC-600 and FBBC-800, corresponding to the 400 $^{\circ}$ C, 600 $^{\circ}$ C and 800 $^{\circ}$ C, respectively. Details about the pyrolysis temper yield of FBBC can be seen in Text S2.

2.3. Characterization

The structure and morphology of FBBC were observed by scanning electronic microscopy (SEM, Nova NanoSEM230) and high-resolution transmission electron microscopy (HRTEM, Tecnai G2 F20). The pore size distribution and specific surface area were measured by the Quantachrome NovaWin (NOVA 2000e) and the degassing conditions are 300 °C for 4 h. The surface functional groups were

investigated by a Fourier-transform infrared spectroscopy (FTIR). Raman spectra were investigated at the excitation wavelength of 532 nm, accompanied with the range from 100 to 4000 nm. X-ray photoelectron spectroscopy (XPS) was conducted on Thermo Escalab 250XI. Zeta potential was obtained from the Zetasizer Nano (Zetasizer Nano ZSP). Electrochemical measurements details including electrochemical impedance spectroscopy (EIS), liner sweep voltammetry (LSV) and chronoamperometric measurements were appended in Text S3.

2.4. Catalytic tests and active species detection

The catalytic oxidation experiment of phenol was concerned in a thermostatic oscillator (170 rpm, 25°C). Unless otherwise states 3 mg FBBC and 30 mg PDS were added into 30 mL of phenol solution (20°pm) to initiate the degradation reaction. At specific time intervals, 1 mL of samples was withdrawn, filtered by 0.45 µm membrane and deterted by phenol concentration immediately. The adsorption experiment was terformed in the same conditions without the addition of PDS. The experimental parameter selection was provided in Text S4. Phenol concentration was analyzed using a high-performance liquid chromatography (acetonitrile/water=30/70, volume ratio) at the wavelength of 270 nm. All the experiments were conducted in triplicate.

Pseudo-first order kinetic model with high values of R^2 was used to mimic the phenol degradation curves in this study. The reaction rate constant k_{obs} was calculated by Eq. (1) and Eq. (2):

$$-dC/dt = k_{obs}C$$
 Eq. (1)

 $\ln (C_t/C_0) = -k_{obs}t$ Eq. (2)

 C_t is the concentration of phenol at reaction time t, and C_0 is the initial concentration of phenol.

To identify the roles of reactive radicals in the degradation process, different molar ratios of radical scavengers (methanol (MeOH), tert-butyl-alcohol (TBA) and ethanol (EtOH)) were added into the reaction system for the quenching of SO₄ · and OH. Besides, 10 mM of p-benzoquinone (BQ), NaN₃ and K₂CrO₄ were used to scavenge superoxide radical (O₂ ·), singlet oxygen (¹O₂) and free electrons, respectively. An electron spin resonance spectrometer (Ext. VEOL FA200) with 5,5-dimethyl-1-pyrroline N-oxide (DMPO) and 2 · 6,6-tetramethyl-4-piperidinol (TMP) as capture agent was also used to deterning the generated active radicals.

177 3. Result and Discussion

3.1. Morphological characterization of IBBC

The morphology and fruction characterization of FBBC are investigated by SEM and TEM. It is observed from Fig. 1a-b that FBBC-800 exhibits rough, wrinkle-rich surface accompanied with high specific area of 758.44 m²/g (Table S1). It is estimated that the high specific area of FBBC-800 is caused by the nanohydroxyapatite self-template, which is inserted into the carbon frame to generate micro-pores during pyrolysis, and then can be mostly removed from the product by diluted acid solution (Medellin-Castillo et al., 2014). As can be seen from Fig 1c, some spiral stripe-like nanosheet are observed in FBBC-800, which could be the stacked graphite layers formed by strong π - π interactions. TEM

mapping shows the existence of C, O and N, among which the N element may be derived from proteins and fats (Fig. 1d-f). The average pore size of FBBC-800 is determined to be 1.17 nm, showing the micro-pore structure (Table S1). The higher specific area and abundant micro-pore structure of FBBC-800 would provide more active site for the catalytic reaction. Besides, the porosity of biochar is largely affected by pyrolysis temperature. As shown in Fig. S1, FBBC-400 presents a relative blocked structure. With the increase of pyrolysis temperature, the structures of solid biochar become condensed and more volatile organic ma (e.g. bio-oil) and gases (e.g. CO₂, CH₄, CO, H₂) are generated (Kambo and 3, 2015). The loss of those volatile compounds contributes to the leveloped pores in biochar. Moreover, at higher pyrolysis temperature, the parolysis process transitioned from the liquification reactions (generation of bio ii) to the release of gasses, and further (Eq. (3-6)) (Bommier et al., 2015; Niu et went through some self-activation al. 2019b). Therefore, higher pyrolysis temperature leads to a significant The specific area of FBBC is in the order of FBBC-400 enhancement of $(0.48 \text{ m}^2/\text{g}) < \text{FBBC-600} (343.41 \text{ m}^2/\text{g}) < \text{FBBC-800} (758.44 \text{ m}^2/\text{g}) \text{ (Table S1)},$ which is in line with the results of SEM and TEM images (Fig. S1).

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$$C+ H_2O \rightarrow CO+ H_2$$
 Eq. (3)

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$$C + CO_2 \rightarrow 2CO$$
 Eq. (4)

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$$C + 2H_2 \rightarrow CH_4$$
 Eq. (5)

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$$2Ca_6(PO_4)_3(CO_3)(OH) \rightarrow 2Ca_{6-\gamma}(PO_4)_3 (CO_3)_{1-\gamma}O_{\chi/2} (OH)_{1-\chi} + 2\gamma CaO +$$

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$$\chi H_2O + 2\gamma CO_2$$
 Eq. (6)

3.2. Catalytic degradation performance

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As the structure of biochar is largely affected by the pyrolysis temperature, three kinds of FBBC prepared at 400 °C, 600 °C and 800 °C are studied and their catalytic performance is evaluated by the degradation of phenol (a model pollutant). As shown in Fig. 2a, adsorption contributes a little to the removal of phenol. Among all the biochar samples, FBBC-800 presents the highest adsorption capacity, with 10.7% of phenol removal within 60 min, followed by the order of FBBC-600 (5.5%) and FBBC-400 (1.2%). This result proves the low potential of FBP as adsorbents for direct application (Naushad et al., 2019a). The higher portion capacity of FBBC-800 was probably ascribed to the larger spear Sic surface area $(758.44 \text{ m}^2/\text{g})$ and strong π - π interaction (Senthil Kumar oll). Chemical oxidation of phenol by PDS was displayed in Fig. 2b. Neably, PDS alone could hardly degrade FBBC (0.1 g/L) and PDS (1 g/L), the phenol. However, in the preser removal efficiency of phanol has significantly improved, with the order of FBBC-400. FBBC-800 exhibits the best catalytic FBBC-800> FB performance of 100% phenol removal in 60 min. Therefore, FBBC-800 was selected as the representative catalyst for the subsequent research. The degradation efficiency was comparable or even better than other metal-free catalyst (Table S2). More importantly, this material was much easier to prepare and cost effective. As the concentration of PDS rising from 0.5 to 2 g/L, the rate constant k_{obs} increased from 0.06119 to 0.10886 min⁻¹, due to more reactive radicals produced with higher PDS amount (Fig. 2c). However, further increasing the PDS dosage to 3

g/L resulted in a minor decrease in k_{obs} , which could be explained by the self-quenching of excessive radicals (Tang et al., 2018). The effect of FBBC dosage was studied in Fig. 2d. With the increase of FBBC dosage from 0.33 to 1.33 g/L, an enhanced removal of phenol was achieved, along with the rate constant k_{obs} from 0.01082 to 0.11827 min⁻¹. Higher dosage of FBBC provides more catalytic site for both adsorption and oxidation, and increases the effective collision between catalyst and pollutant, contributing to better removal capacity for phenol (Wang et al. 2019a). Taken together the reaction rates and reaction cost, 1 g/L PLS ard 0.1g/L FBBC was selected for the subsequent experiment.

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The initial pH of the solution is another fact that influences the catalytic capacity of FBBC (Qin et al. 2019). As is sh w ng. 2e, no obvious change in phenol degradation was observed with the Hranging from 3 to 9, indicating the em for practical water treatment. Besides, wide applicability of FBBC-800 a slight enhancement in de rada on can be seen as the increase of initial pH, with 603 min⁻¹ at the pH of 9. Duan et al. (2016a) reported that the maxima k_{obs} OH was easier to lonate an electron to PDS than H₂O molecules to produce hydroxyl radicals, which might be the reason for the facilitated PDS activation at pH=9. However, at strong alkaline condition (pH=11), the removal efficiency of phenol sharply declined (59%), along with the k_{obs} down to 0.01352 min⁻¹. The electrostatic repulsion between phenol and FBBC-800 was responsible for the decrease in phenol removal (Dhiman et al., 2017; Kumar et al., 2018). The surface of FBBC-800 was negatively charged at pH >3 (Fig. S2). In addition, the pK_a of phenol was 9.95, indicating that phenol existed in the form of $C_6H_5O^-$ at pH > 9.95 (Sun et al., 2020). Therefore, the interaction between FBBC-800 and phenol was not favored.

The influence of initial phenol concentration was also investigated and presented in Fig. 2f. It was clear that 100% of phenol removal was achieved at phenol concentration of 10 mg/L, and 70.8% removal was still obtained at higher phenol concentration of 40 mg/L, which manifested the good performance of FBBC-800/PDS system under different concentrations. However the rate constant k_{obs} decreased from 0.1865 to 0.01864 min⁻¹ with the increasing of initial phenol concentration from 10 to 40 mg/L. This could be that high pollutant concentration resulted in the over-occupied active sites and recreased the catalytic activity of FBBC-800 for PDS activation (Alshehri et al. 2014).

3.3. Identification of reactive sees

Until now, both radical routes and non-radical routes have been proved in catalyst/PDS system. To verify the involvement of radical in this reaction, radical quenching experiments were carried out (Song et al., 2019). Firstly, MeOH (k_{SO4} – and k_{OH} are 1.1×10^7 M⁻¹ s⁻¹ and 9.7×10^8 M⁻¹ s⁻¹, respectively) and TBA (k_{SO4} – and k_{OH} are 4×10^5 M⁻¹ s⁻¹ and 6×10^8 M⁻¹ s⁻¹, respectively) were used as radicals scavengers for quenching the OH and SO₄ · (Buxton et al., 1988; Neta et al., 1988). As presented in Fig. 3a-b, both MeOH and TBA have suppressed phenol degradation. Addition of MeOH resulted in an insignificant decrease of phenol degradation (83.28% removal), even with high dose [MeOH /PDS=1000].

Surprisingly, TBA at low amount [TBA /PDS=100] brought minor inhibition effect (93.13% removal) on phenol degradation, but exhibited significant quenching effect (34.94% removal) at high amount [TBA /PDS=1000]. Generally, MeOH was supposed to have comparable or even better inhibition effect in PDS than TBA, as MeOH was reactive with both SO₄ • and OH but TBA had higher reactive rate with OH. The contradictory phenomena in this study may be related to the following two aspects: 1) Although one molecule MeOH could react with one molecule SO₄ -, the resultant CH₂OH ·could again react with POS produce SO₄ • and directly involve in phenol degradation (Yu et al., onsequently, the inhibition effect of MeOH is alleviated, which is consistent with the previous study (Bao et al., 2018). 2) TBA had stronger (ff) at with FBBC-800 due to the hydrophobic interaction, and impeded the access of PDS and phenol to FBBC-800. orted to exhibit high rate for both SO₄. In addition, ethanol (EtOH) w and OH (k_{SO4} – and k_{OH} are $10^7 M^{-1}$ s⁻¹ and $10^9 M^{-1}$ s⁻¹, respectively) (Sun et al. ewed the only about 68.1% phenol was removed with high 2020). Fig. 3c 🔊 concentration of EtcH. Totally, the radical quenching experiment indicated that both SO₄ • and OH participated in the reaction. However, the remained high removal of phenol even when radicals were captured implied the possible existence of other reactive species. Then, BQ was used to confirm the involvement of O₂. $(k_{02} = 1 \times 10^9 \text{ M}^{-1} \text{ s}^{-1})$ (Naushad et al., 2019b; Rao and Hayon, 1975). As presented in Fig. 3d, BQ moderately suppressed phenol removal, verifying the involvement of O₂ · in phenol degradation. ¹O₂, as a reactive oxygen species derived from

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non-radical pathway, was confirmed by quencher NaN₃ (Oh et al., 2018; Zhu et al., 2018). An evident inhibiting effect was observed after addition of NaN₃, with only about 42% phenol removal (Fig. 3d), indicating that ${}^{1}O_{2}$ played an important role in this reaction.

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To further identify the existence of radicals in FBBC-800/PDS system, ESR spectroscopy was conducted with DMPO and TMP as spin trapping agents (Fig. 4). When PDS was used alone, no radical signal peak was detected. However, when PDS and FBBC-800 was simultaneously added, the signal peak of PMPO-OH (four lines in 1:2:2:1; $\alpha_N=\alpha_H=14.9$ G) and DMPO-SO₄ (six line $1:1:1:1:1:\alpha_N=13.2$ G, $\alpha_{v-H}=1.48$ G, $\alpha_{v-H}=0.78$ G, $\alpha_{B-H}=9.6$ G,) were obviously observed and DMPO-OH had higher relative intensity than DMPOg. 4a). This confirmed the existence of SO₄ · and OH, and a higher arount of OH than SO₄ ·. Besides, as d of the signal intensities was observed, the reaction proceeded, an incre roduction of radicals. In addition, four characteristic implying the continuous eloging to O_2 • were also found in Fig. 4b. When TEMP peaks of DMPO was added, triple signal peak with an equal height signal (three lines in 1:1:1, splitting constants of $\alpha_N=16.9$ G) was obviously seen (Fig. 4b). Combining with the radical quenching experiments and ESR analysis, it can be inferred that $SO_4 \cdot , OH$, O₂ - and ¹O₂ all take part in the reaction, among which OH and ¹O₂ play a dominate role.

3.4. The structure changes and surface investigation

The morphological changes of FBBC-800 before and after reaction were

investigated by SEM, which showed no significant difference (Fig. S4). Besides, the porosity is also important for a catalyst. It is well known that porous structure facilitates adsorption of pollutant and contributes to more exposed active sites for the reactions (Wang et al. 2019b). Ho et al. (2019) found that spirulina residue pyrolyzed at 900 °C showed 100% sulfamethoxazole removal in PDS system while the desalted spirulina residue pyrolyzed at 900 °C displayed only 5.2% sulfamethoxazole removal, due to the extremely lower surface area of the latter than the former. In this study, FBBC-800 without HCl treatment hall specific area (96.752 m²/g) than FBBC-800 (758.44 m²/g) (Table S1), anied with about 57.2% phenol removal (Fig. S3a), which indicated the importance of catalyst porosity in PDS system. Consistently, the higher specime area of FBBC-800 (758.44 m^2/g) than FBBC-600 (343.41 m^2/g) and FBBC-400 (0.48 m^2/g) may partially explain its higher catalytic To examine the porosity change of FBBC-800 after reaction, N adserption-desorption isotherms was performed, which The isotherm curve as presented in Fig. 5a is a typical type was shown in Fig. I curve, showing the characteristic of micro-pore, which is consistent with the result of pore size distributions curves. Compared with fresh FBBC-800, the specific surface area and total pore volume of used FBBC-800 reduced by 19.7% and 16.3%, respectively. The decline in specific area after reaction may be ascribed to adsorption of intermediates, and the consequently blocked active site. However, in our study, the specific surface area of used FBBC-800 is not significantly decreased relative to fresh FBBC-800, implying that blocked porosity may not be the main

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reason for the deterioration of catalyst (Fig. S3b).

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Moreover, the surface change of FBBC-800 after reaction was investigated. Many kinds of oxygen-containing groups have been reported to influence the catalytic activity of carbon-based catalyst, including carboxyl (O=C-OH), carbonyl (C=O) and hydroxyl (C-OH). Duan et al. (2016b) conducted a theoretical calculation and identified the electron-rich ketonic group (C=O) as the primary active site for peroxymonosulfate activation among the oxygen-containing groups. The oxygen atom with lone-pair electrons in ketonic group ares high affinity with peroxymonosulfate and promote the electron tra rocess. Fig. S5 presented the FTIR of FBBC-800 before and after action. The band around 3389 cm⁻¹ was attributed to the -OH stretching vib atton and 2859-2925 cm⁻¹ was derived from aliphatic C-H stretching vibration (Zhao et al. 2016). The distinct peak c C=C stretching (Tang et al. 2018). The at 1584 cm⁻¹ might belong to the band for C=O stretching at about 1696 cm⁻¹ was also observed (Zhao et al. 2016). 11.3 cm⁻¹ was ascribed to C-OH vibration (Zhang and In addition, the \square Wang. 2016). After reaction, the intensity for –OH vibration increased and for C=O vibration decreased, which suggested the possible conversion of the two groups during the catalytic reaction. For further analysis, XPS survey of FBBC-800 was conducted (Fig. 5c). It was apparent to see three distinct peaks at around 285.19 eV, 400.19 eV and 532.19 eV, corresponding to C1s, N1s, and O1s. Clearly, after reaction, the O content in FBBC-800 increased from 9.18% to 13.10%, which revealed that the surface of FBBC-800 was oxidized during PDS activation. This

can be explained in two ways (Tang et al., 2018). On the one hand, surface functional groups act as electron transfer intermediate and are oxidized by providing electrons to activate PDS. On the other hand, the production of abundant radicals leads to a highly oxidative environment around FBBC-800, eventually inducing the surface oxidation. The O1s spectra of the fresh FBBC-800 could be fitted into three main peaks at 530.51, 531.31 and 532.84 eV, assigning to O=C-OH, C=O and C-OH, respectively. It was observed that after reaction both the content of O=C-OH and C=O decreased, from 11.5% to 3.6% and 33.5% respectively, accompanied with the increase of C-OH (from 55% to 73) 6a and c). This indicated the conversion of C=O to C-OH during the catalytic process. In addition, nitrogen doping is an efficient way to improve he reactivity of carbon-based catalyst. The main peak of N1s around 39836 eV and 400.37 eV was assigned to et al. (2019) reported that graphitic N had pyridnic N and pyrrolic N (Fig. PMS and thus promote PMS activation. Wang et al. high adsorption energy with e p trogen configurations such as pyrrolic N and pyridinic (2019a) claimed N might be the active site for PDS activation. However, the close ratio of pyridnic N and pyrrolic N before (20%:80%) and after (17.36%:82.64%) reaction indicates that nitrogen element may not act as the catalytic site for PDS activation in this study (Fig. 6b and d). The decline in the relative content of N was likely due to the introduced exogenous oxygen rather than the catalytic loss.

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The defective edge of carbon materials has also been identified as catalytic sites for PDS oxidation. Raman spectra, a powerful technique for exploring the

crystalline structure of carbon materials, were conducted and the related parameter I_D/I_G (an indicator of disorder degree) was calculated. As shown in Fig. 5d, two important characteristics peaks appeared at FBBC-800 both before and after reaction, where the D peak (1350 cm⁻¹) represented the disorder degree and G peak (1600 cm⁻¹) reflected the graphitization level of carbon materials. Compared to original, the I_D/I_G of FBBC-800 after reaction decreased from 0.972 to 0.945, indicating the decrease of disorder degree. This was probably caused by the break of defective edge, alteration of surface functional group and the also ration of organic matters on sp² hybridized carbon (Tang et al. 2018). Based on above studies, we suppose that the reduced defective sites in FBBC81 used may be responsible for the deterioration of catalytic performance.

3.5 Electron transfer pathway

Based on above analysis, the coarse mechanism of phenol in FBBC-800/PDS system is proposed and precented in Scheme 1. For the radical pathway, it has been proved that SO₄ C. Oheano O_2 participate in the reaction. Since $S_2O_8^{2-}$ itself is incapable of generating OH, it must be derived from H_2O or OH^- (Howard Jr and Levitt, 1953). This process should be assisted by external energy, such as UV, thermal, ultrasound and catalyst. Previous studies demonstrated the sp^2 carbons, defective edges and ketone groups were chemically active to stimulate persulfate decomposition and generate radicals (Duan et al., 2016a; Duan et al., 2016b). The electron-rich regions (e.g. the sp^2 -carbon with defective edge and ketonic groups) on FBBC-800 could affect the electronic configuration of PDS (especially the O-O

bond in $S_2O_8^{2-}$) to form the "semi-active" $S_2O_8^{2-}$ complex. In this case, the hydrolysis reaction of PDS was activated (Eq. (7)). Then the produced HO₂⁻ would react with another $S_2O_8^{2-}$ quickly and the SO_4^{-} and O_2^{-} was generated (Eq. (8)). Meanwhile, $SO_4 \cdot could$ quickly transform into OH through Eq. (9) ($k = 7 \times 10^7 \text{ M}^{-1}$ s⁻¹) (Liu et al., 2016), which might be the reason for the strong OH signals and weak SO₄ • signals in ESR. As to ¹O₂, an important species in the non-radical pathway, the detailed generation mechanism has not been fully understood. A possible pathway could be through the reaction of O_2 $\bar{\cdot}$ and H_1O_2 ed on Eq. (10) and the C=O group played an important role (Cheng et al.,

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$$S_2O_8^{2-} + 2H_2O \rightarrow HO_2^{-} + 2SO_4^{2-} + 3H^+$$
 Eq. (7)

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$$S_2O_8^{2-} + HO_2^{-} \rightarrow SO_4^{2-} + SO_4^{-} + O_2^{-} + O_2^{-}$$
 Eq. (8)

419
$$SO_4 \cdot + OH^- \rightarrow SO_4^{2-} + OH$$
 Eq. (9)

420
$$2O_2 \cdot + 2H_2O \rightarrow H_2O_2 + 2O_1 \cdot + O_2$$
 Eq. (10)

421 FBBC + PDS
$$\rightarrow$$
 [FB C-PDS*] complex Eq. (11)

422
$$S_2O_8^{2-} + 2e \longrightarrow 2SO_4^{2-}$$
 Eq. (12)

As mentioned bove, the formation of charge transfer intermediates between PDS and FBBC-800 accelerate the decomposition of PDS and initiate the subsequent radical generation reaction for phenol degradation. Besides, the direct electron transfer in the non-radical pathway also contributes to the degradation of phenol. When both PDS and phenol adhere to the FBBC-800, the ternary system is formed, where the electron can be abstracted from adsorbed phenol (electron donor) to metastable FBBC-PDS* complexes (electron receptor), with FBBC-800 as

electron medium (Eq. (11)) (Ren et al. 2019). The O-O bond in PDS is cleaved by accepting the electron and finally decomposed into SO_4^{2-} (Eq. (12)). To prove the point, K₂CrO₄ as an electron trapping agent was added in the PDS system. The inhibited effect as shown in Fig. 3d proved the occurrence of electron transfer. For further analysis, some electrochemical measurements were conducted. The impedance of carbon-based catalyst is an important parameter affecting the electron transfer, which can be obtained from the Nyquist plots in EIS. The largely decreased semicircle diameter of FBBC-800 indicated its better conductivi than the blank solution with bare GCE (Fig. 7a). Besides, FBBC-800 e. a much smaller semicircle than SSBC-400, manifesting the better enductivity of FBBC-800 (Fig. S6). This may be an important factor for th nigher catalytic efficiency of addit on FBBC-800 than FBBC-400. In can be seen from the rent change was observed after injection chronoamperometry test (Fig. 7 profided direct evidence of electron transfer among of PDS and phenol, which ph nol molecules. The LSV curves (Fig. 7c) also FBBC-800, PD and demonstrated the increased current after addition of PDS, suggesting the formation of metastable complexes between PDS and FBBC-800. Besides, the current was further increased after addition of phenol, which might be ascribed to the electron transfer between phenol, FBBC-800 and PDS. Totally, in the synergistic effect of radical and non-radical pathway, the phenol was completely oxidized. The possible degradation pathway of phenol was provided in Fig. S7.

3.6. Practical application

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The interference of several background substances on the catalytic efficiency of FBBC-800/PDS system was investigated (Yi et al., 2019). As shown in Fig. S8a, the impact of Cl⁻ and HA was insignificant, but HCO₃⁻ and HPO₄²⁻ slightly inhibited phenol degradation. The degradation efficiency decreased from 100% to 74% for HCO₃⁻ and to 80% for HPO₄²⁻. This phenomenon may be ascribed to scavenging effect of HCO₃⁻ and HPO₄²⁻. To investigate the practical efficiency of FBBC-800 in real water matrix, samples taken from Xiangjiang River and Taozi Lake were used for experiment. Efficient removal of phenol was Il obtained in river water and lake water within 60 minutes, which confident the applicability of FBBC-800/PDS system for realistic water environment remediation (Fig. S8b). Besides, the degradation efficiency of 500/PDS to pharmaceutical wastewater was also studied. As the component of pharmaceutical wastewater was of complicated with high various organic compounds, three-dimensional excitation Emission matrix fluorescence spectra was used to organic pollution. Fig. S8c-d showed that the characterize the fluorescence intensity of pharmaceutical wastewater samples had significantly decreased after reaction, indicating the effectiveness of FBBC-800/PDS system in complex water environment. In addition to phenol, the degradation of other three typical pollutants was also conducted, including rhodamine B, tetracycline hydrochloride and 2,4-dichlorophenol. Result in Fig. S8e demonstrated the universality of FBBC-800/PDS system for organic pollutants degradation.

4. Conclusion

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This study developed a low cost and high efficient catalyst by simple pyrolysis of fishbone and used it for persulfate activation. Pyrolysis temperature largely affects the catalytic efficiency of FBBC, with the order of FBBC-800 (100%) > FBBC-600 (65.1%) > FBBC-400 (4.7%). A complete removal of phenol is achieved within 60 min at the FBBC-800 dosage of 0.1 g/L and in a wide pH range (3.0-9.0). Compared with other recently reported metal free catalysts, the FBBC-800 shows competitive or even better catalytic efficiency, simpler preparation process and lower cost. The high surface area, electron transfer capacity, ab and oxygen group and defective structure endow the FBBC-800 with good ca ctivity. Based on the radical quenching experiment, ESR test and electrochemical characterization, both radical and non-radical pathways take pa reaction, and electron transfer Moreover, he FBBC-800/PDS system plays an important role in this process. exhibits good performance in environment, demonstrating its practical tudy provides a new insight into biowaste reutilization application potential. This on f carbon-based catalyst. Further research may keep on and the structure exploitation of the bowaste with hierarchical structure and chemical components for the preparation of highly efficient and reusable catalyst.

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- 729 **CAPTIONS**
- Fig. 1. (a) SEM of FBBC-800; (b, c) TEM of FBBC-800; TEM element mapping of
- FBBC-800: (d) carbon, (e) nitrogen and (f) oxygen.
- 732 **Fig. 2.** (a) Adsorption removal and (b) Oxidation degradation of phenol; (c) Effects
- of PDS dosage; (d) FBBC-800 dosage; (e) Initial pH of solution; (f) Phenol
- concentration. Conditions: [biochar] = 0.1 g/L, [PDS] = 1.0 g/L, [temperature] =
- 735 25 °C, [phenol] = 20 mg/L.
- 736 Fig. 3. Effects of (a) MeOH, (b)TBA and (c) EtOH on plenol degradation at
- different molar ratio with PDS; (d) Effects of NaN₃, B and K₂CrO₄ on phenol
- degradation. Conditions: [FBBC-800] = 0.1 g/L, [NDS] = 1.0 g/L, [temperature] =
- 739 25 °C, [phenol] = 20 mg/L.
- 740 **Fig. 4.** ESR spectra of PDS activation by TSBC-800.
- 741 **Fig. 5.** (a) Nitrogen adsorption and a sorption curves, (b) pore size distribution, (c)
- 742 XPS survey and (d) Raman pectra. FBBC-800 before and after reaction.
- 743 Fig. 6. Ols high resolution scans of (a) FBBC-800 before reaction and (b)
- 744 FBBC-800 after reaction; N1s high resolution scans of (c) FBBC-800 before
- reaction and (d) FBBC-800 after reaction.
- 746 Fig. 7. (a) Electrochemical impedance spectroscopy (EIS) of bare GCE and
- 747 FBBC-800 loaded on GCE; (b) the current–time (I-t) curves of FBBC-800 and (c)
- Linear sweep voltammetry (LSV) curves under different conditions.
- 749 **Scheme 1.** Proposed mechanism of FBBC-800 to activate PDS for phenol
- 750 degradation

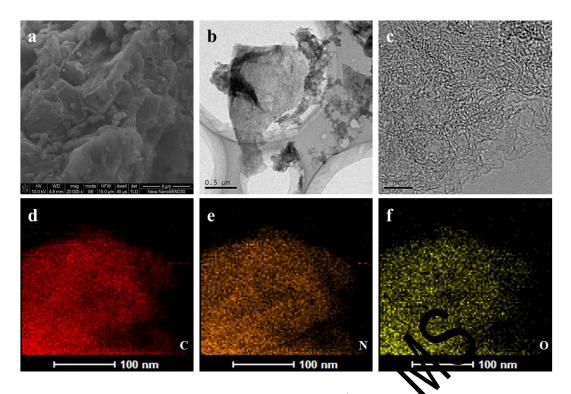


Fig. 1. (a) SEM of FBBC-800; (b, c) TEM of FBBC-800; TEM element mapping of FBBC-800: (d) carbon, (e) nitrogen and (f) oxygen

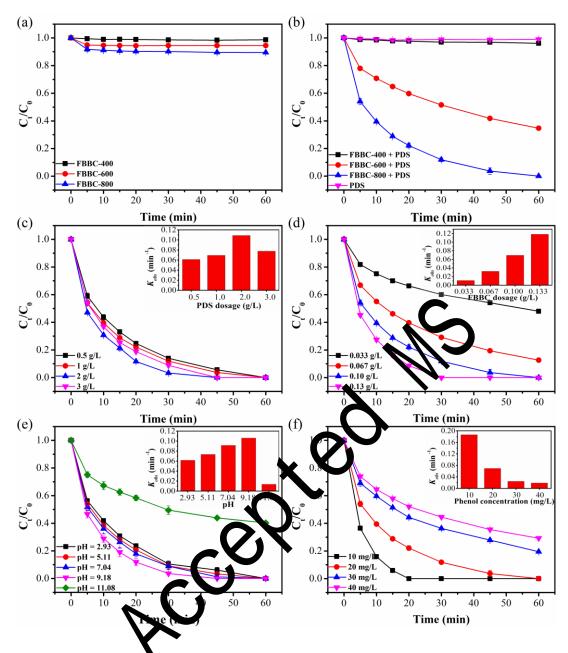


Fig. 2. (a) Adsorption removal and (b) Oxidation degradation of phenol; (c) Effects of PDS dosage; (d) FBBC-800 dosage; (e) Initial pH of solution; (f) Phenol concentration. Conditions: [biochar]=0.1 g/L, [PDS]=1.0 g/L, [temperature]=25 °C, [phenol]=20 mg/L. Insets are the corresponding changes of pseudo-first-order rate constant.

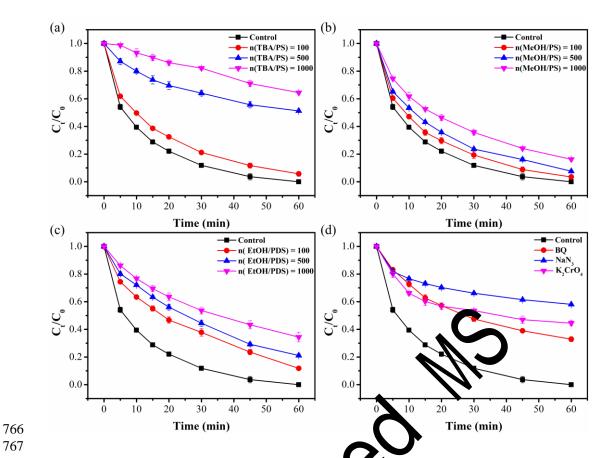


Fig. 3. Effects of (a) MeOH, (b)TBA and OH on phenol degradation at different molar ratio with PDS; (d) Effect of NaN₃, BQ and K₂CrO₄ on phenol g/L, [PDS]=1.0 g/L, [temperature]= $25 \text{ }^{\circ}\text{C}$, degradation. Conditions: [FBBC-800 [phenol]=20 mg/L.

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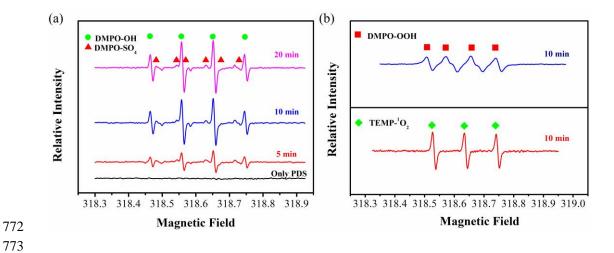


Fig. 4. ESR spectra of PDS activation by FBBC-800.



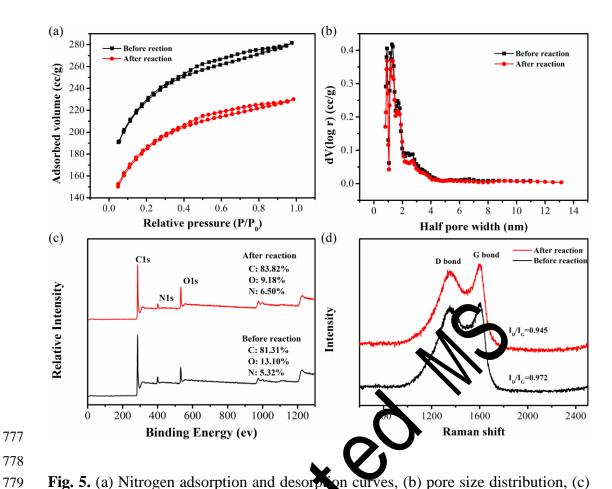


Fig. 5. (a) Nitrogen adsorption and desorption curves, (b) pore size distribution, (c) XPS survey and (d) Raman spectra of A BC-300 before and after reaction.

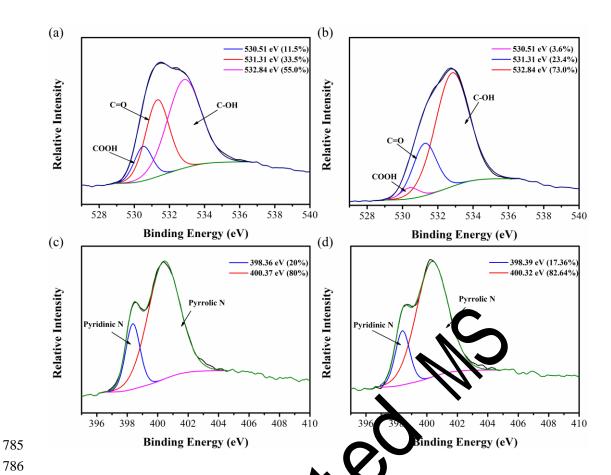


Fig. 6. O1s high resolution scans of (c) FBBC-800 before reaction and (b) FBBC-800 after reaction; N1s high resolution scans of (c) FBBC-800 before reaction and (d) FBBC-800 after reaction.

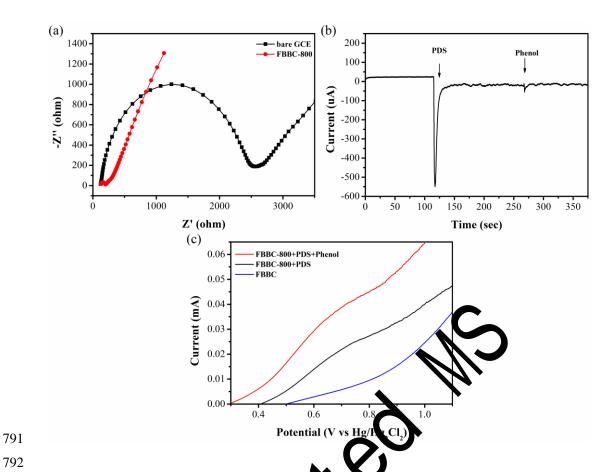
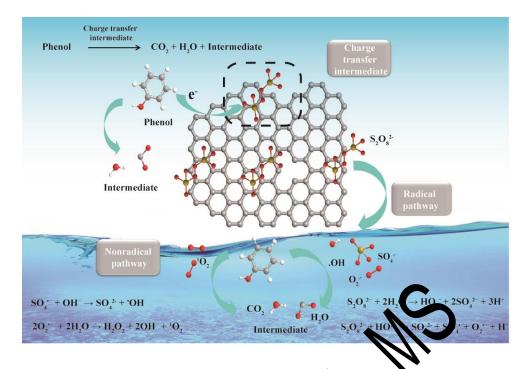


Fig. 7. (a) Electrochemical impedance pectroscopy (EIS) of bare GCE and FBBC-800 loaded on GCE; (b) the carrent—time (I-t) curves of FBBC-800 and (c) Linear sweep voltammetry (LSV) curves under different conditions.



Scheme 1. Proposed mechanism of FBBC-800 to activate PDS for phenol degradation

Abbreviation	Full name
FBBC	Fishbone derived biochar
k_{obs}	Reaction rate constant
PS-AOPs	Persulfate-induced advanced oxidation processes
PDS	Peroxydisulfate
ОН	Hydroxyl radicals
SO_4 -	Sulfate radicals
O_2 $ar{ullet}$	Superoxide radical
$^{1}\mathrm{O}_{2}$	Singlet oxygen
ESR	Electron spin resonance spectromete
FTIR	Fourier-transform infrared spectroscopy
SEM	Scanning electronic nicroscopy
HRTEM	High-resolution transmission electron microscopy
FTIR	Fourier-transform imrared spectroscopy
XPS	X-rey photoelectron spectroscopy
EIS	Electric chemical impedance spectroscopy
LSV	Liner sweep voltammetry
МеОН	Methanol
TBA	Tert-butyl-alcohol
EtOH	Ethanol
BQ	p-benzoquinone
DMPO	5,5-dimethyl-1-pyrroline N-oxide
TMP	2,2,6,6-tetramethyl-4-piperidinol